

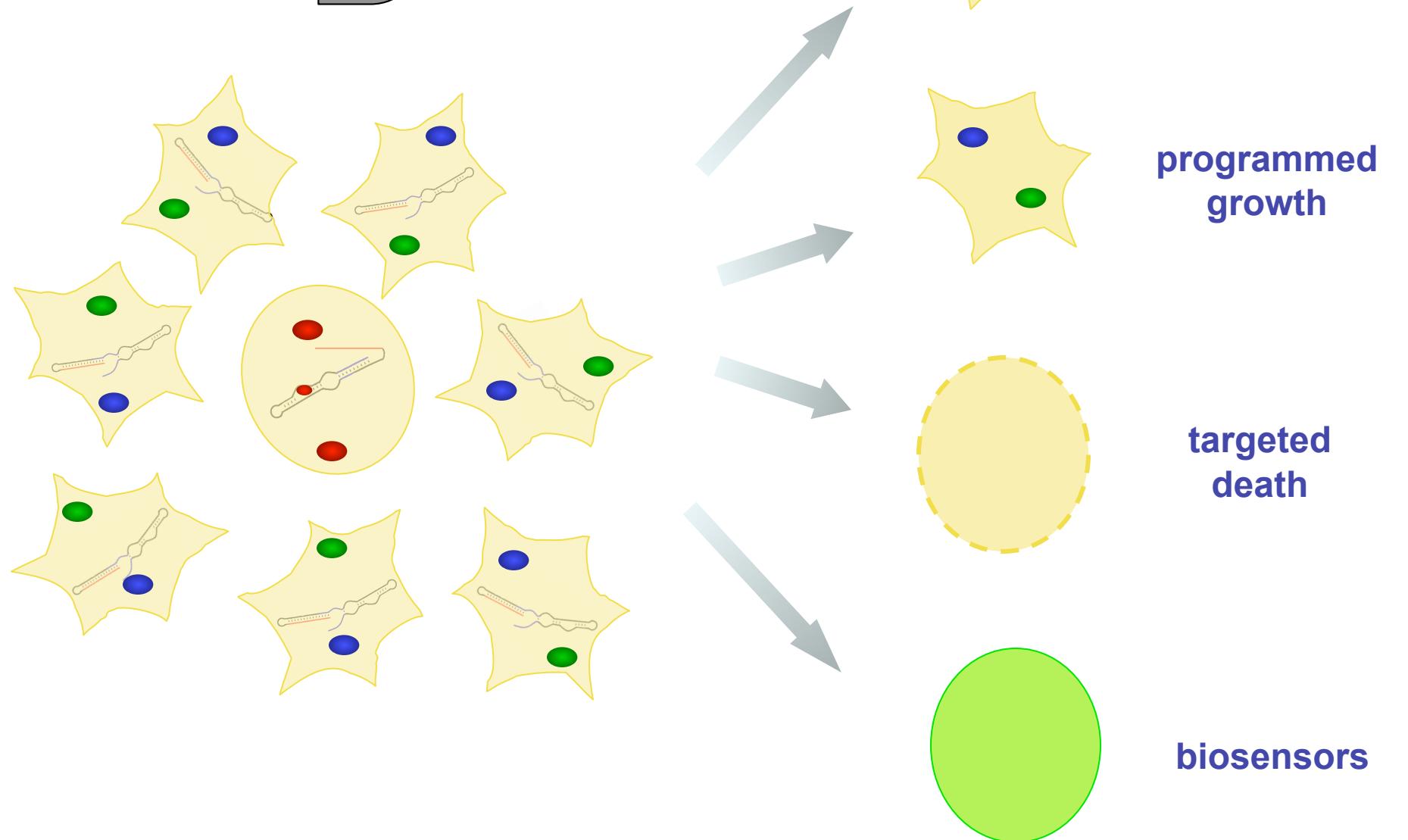
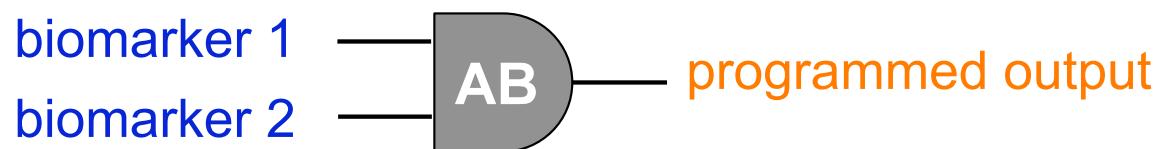
# Programming cellular behavior with RNA controllers

Christina D. Smolke  
Department of Bioengineering  
Stanford University

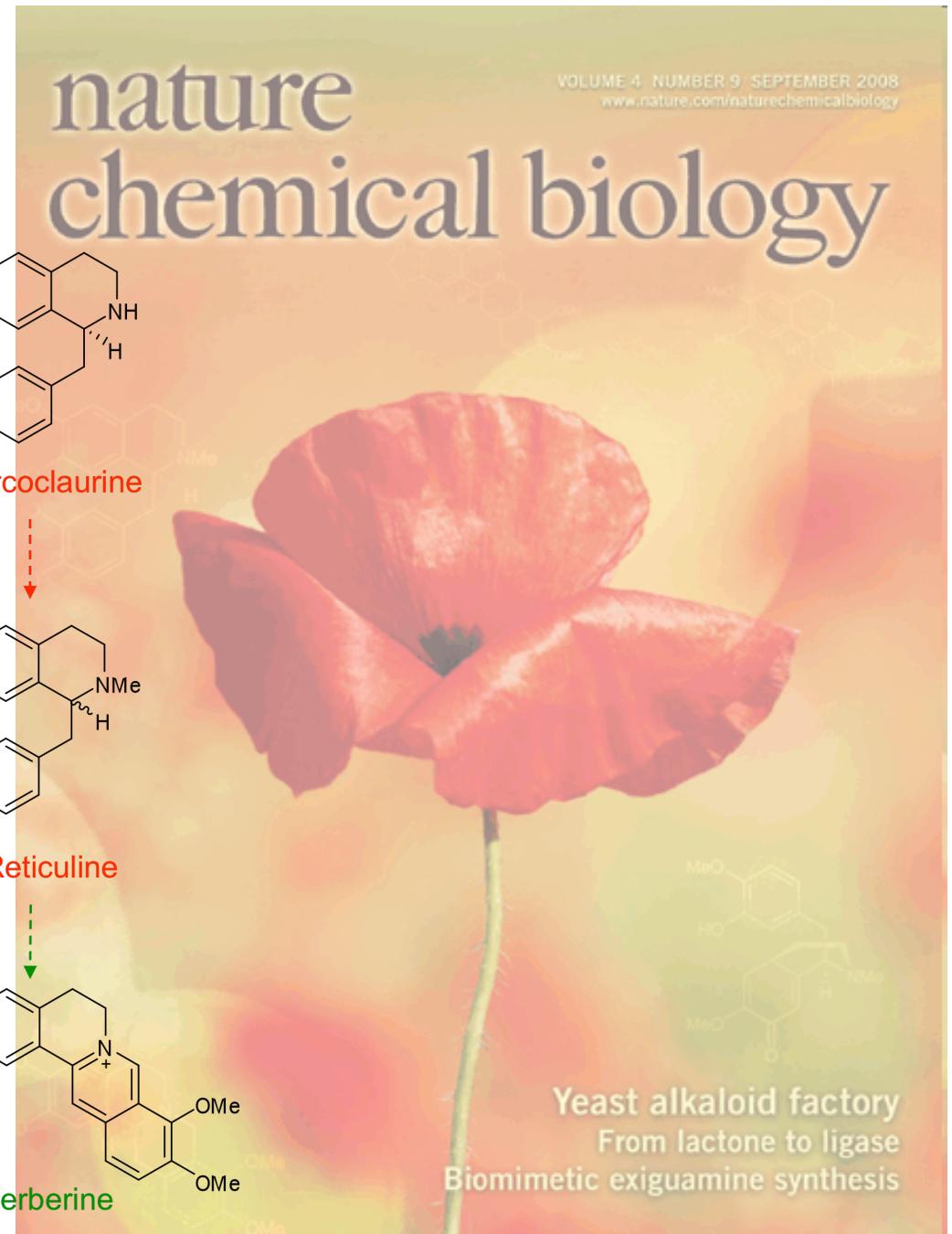
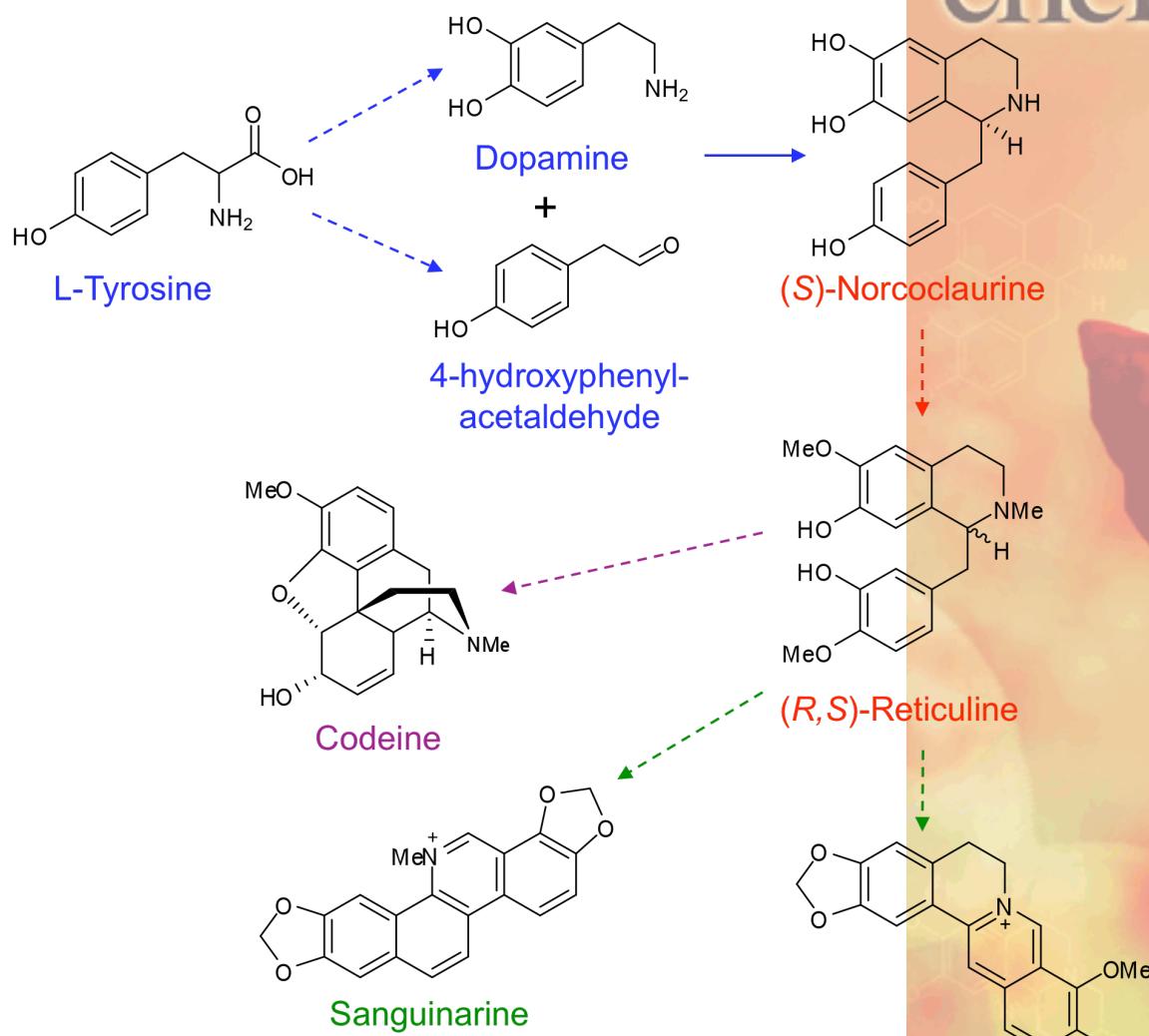
July 13, 2010  
RoSBNet Synthetic Biology Workshop  
St. Anne's College, University of Oxford



# Targeted therapeutics



# Manufacturing and discovery platforms



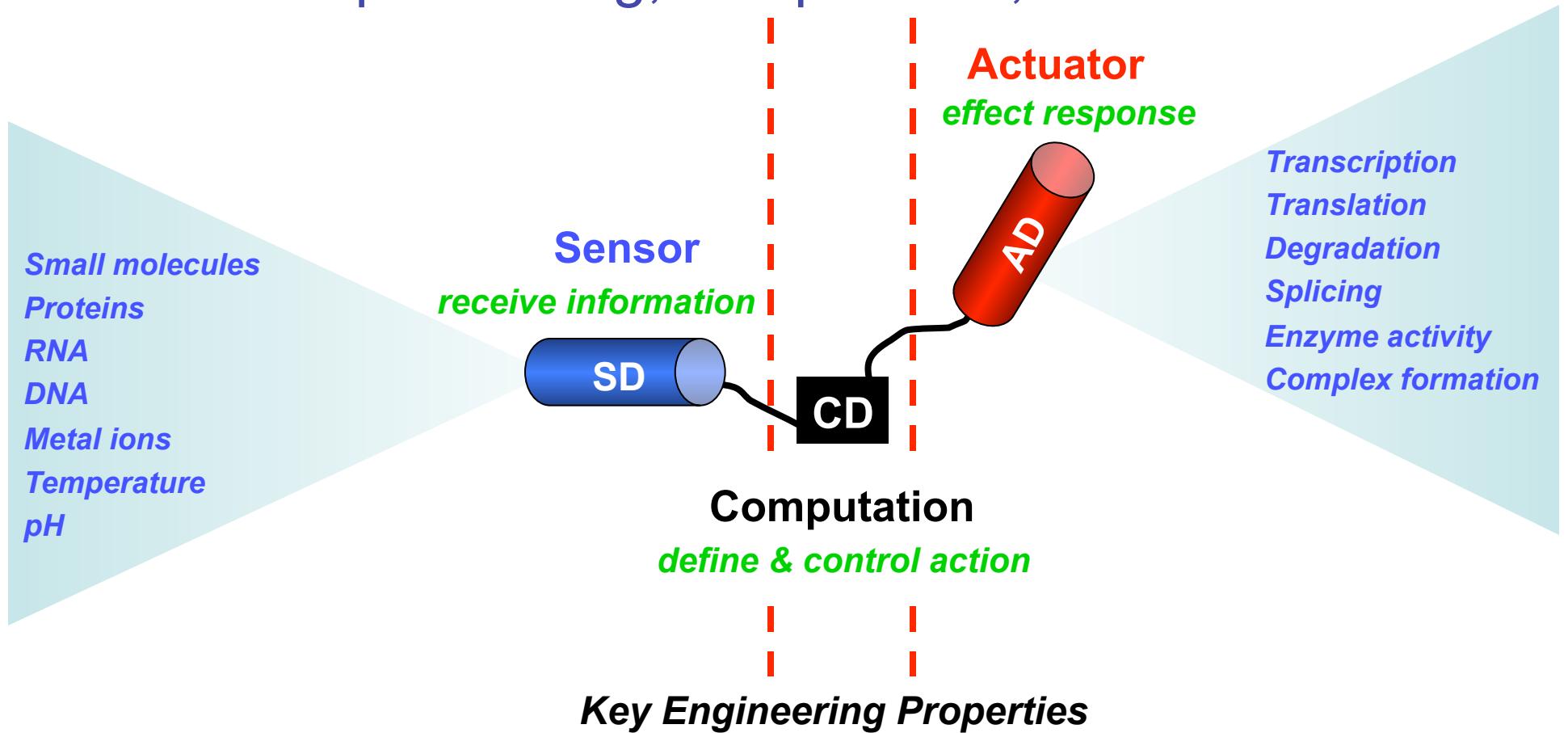
Hawkins KM, Smolke CD. 2008. *Nat Chem Biol.* 4:564-73

# Apps

# Tools

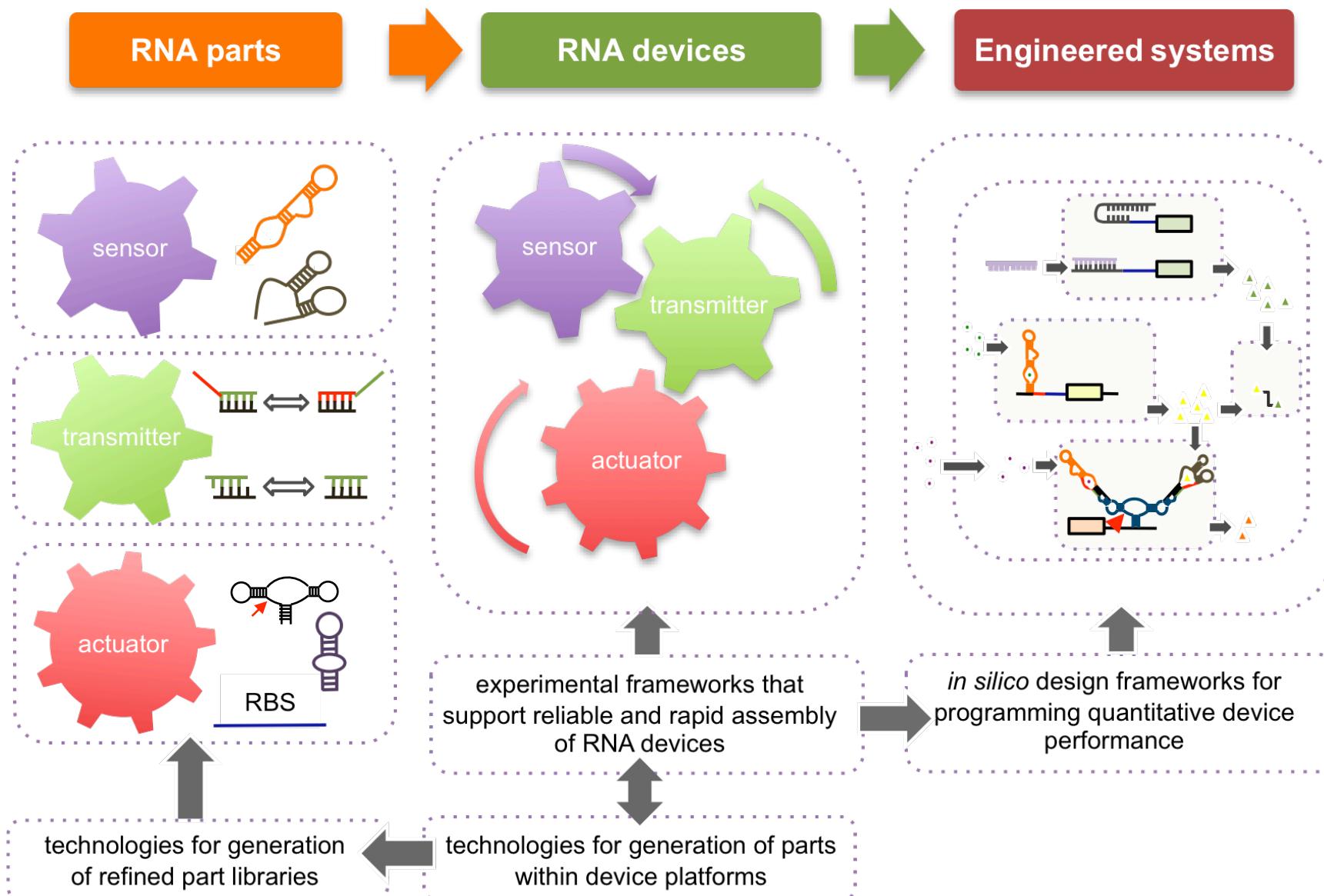
1. Info. theory & signal proc.
2. Device design
3. Languages & grammars
4. Standards & abstraction
5. Fab, CAD & EDA
6. Reverse engineering
7. Control & dyn. systems

# Information processing, computation, and control

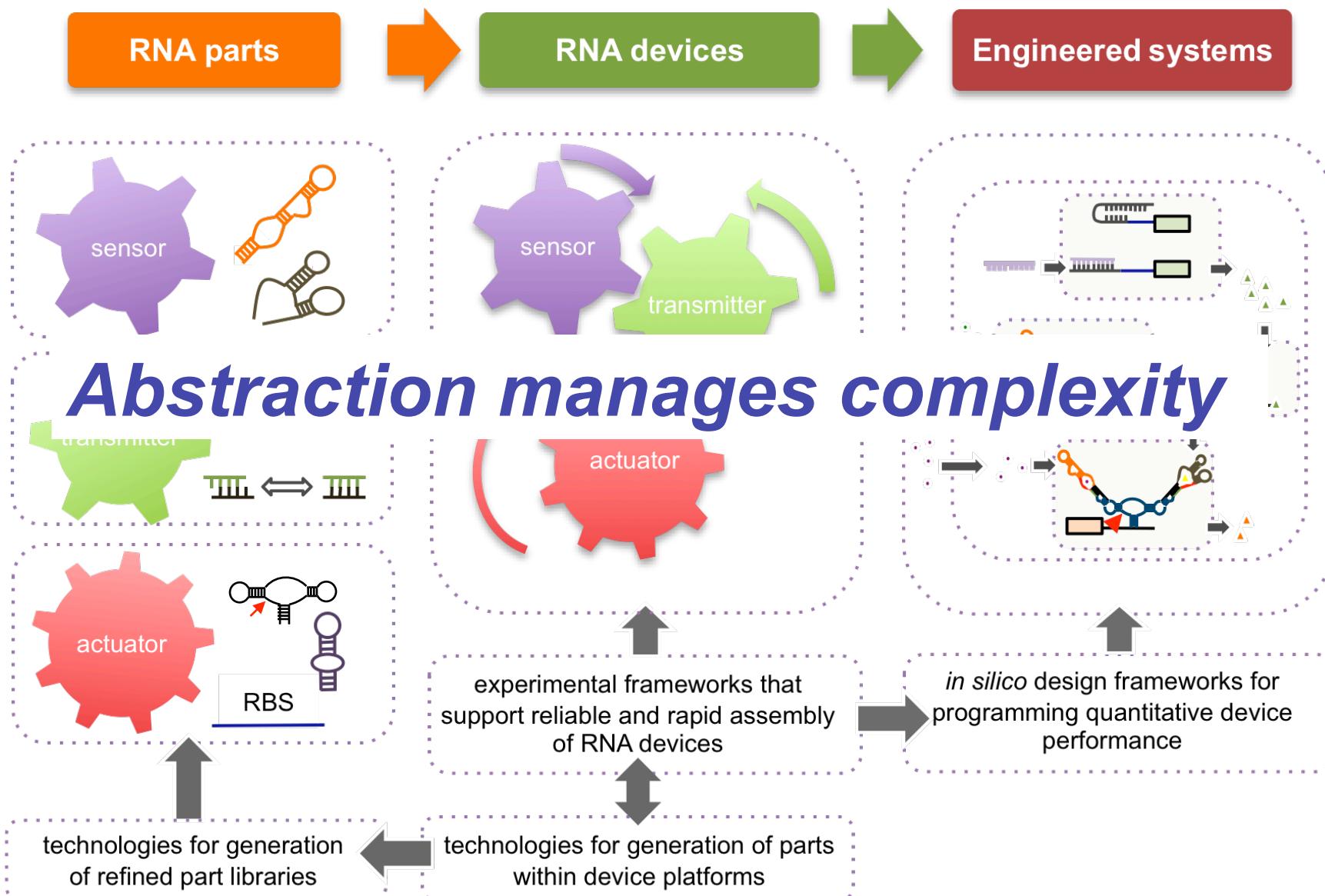


- ✓ scalability
- ✓ portability
- ✓ utility
- ✓ composability
- ✓ reliability

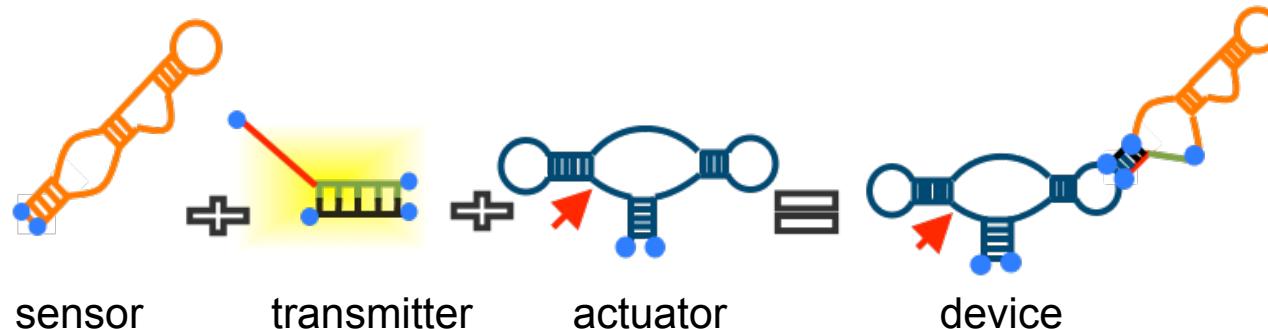
# Plan for developing a programmable input/output tool



# Plan for developing a programmable input/output tool

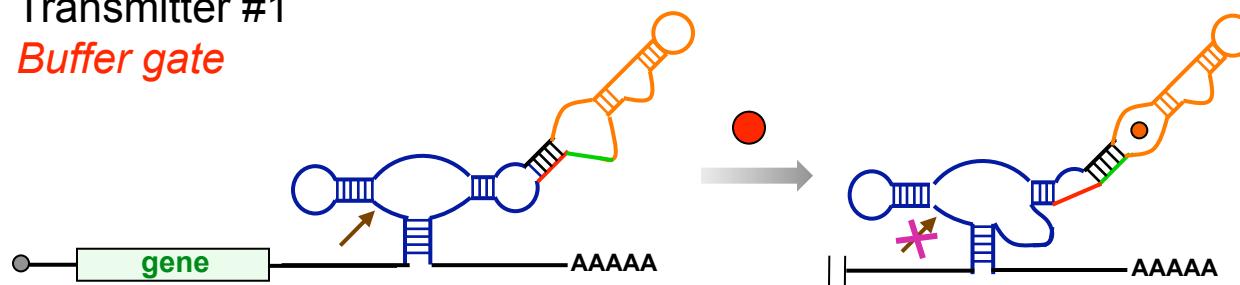


# A platform for programming device function

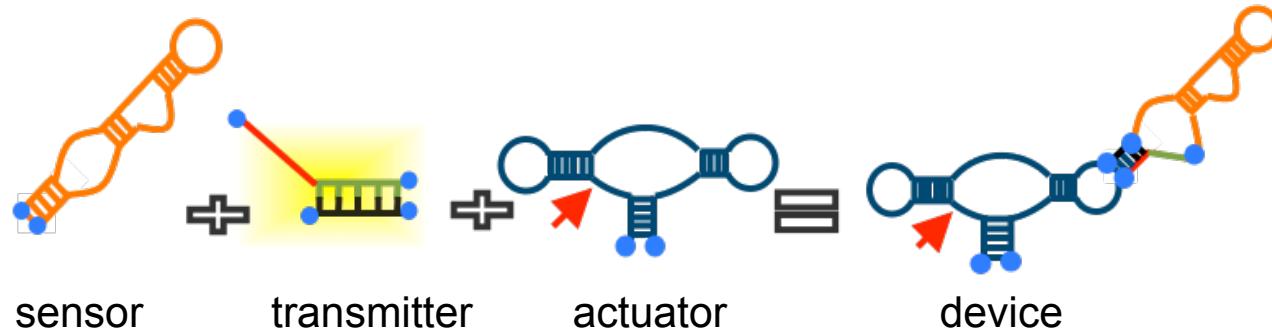


Transmitter #1

*Buffer gate*

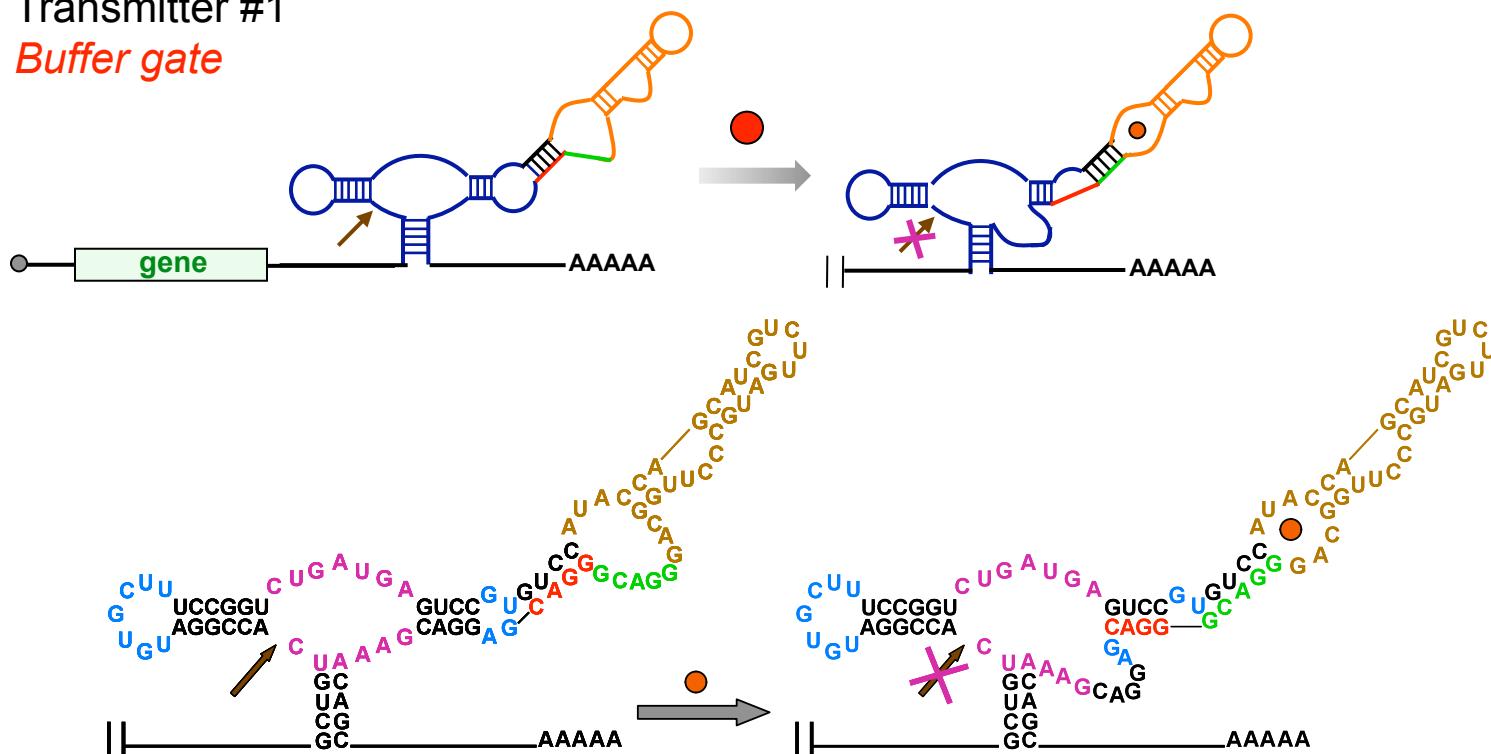


# A platform for programming device function

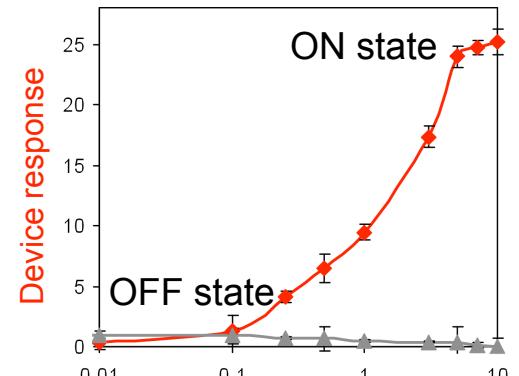
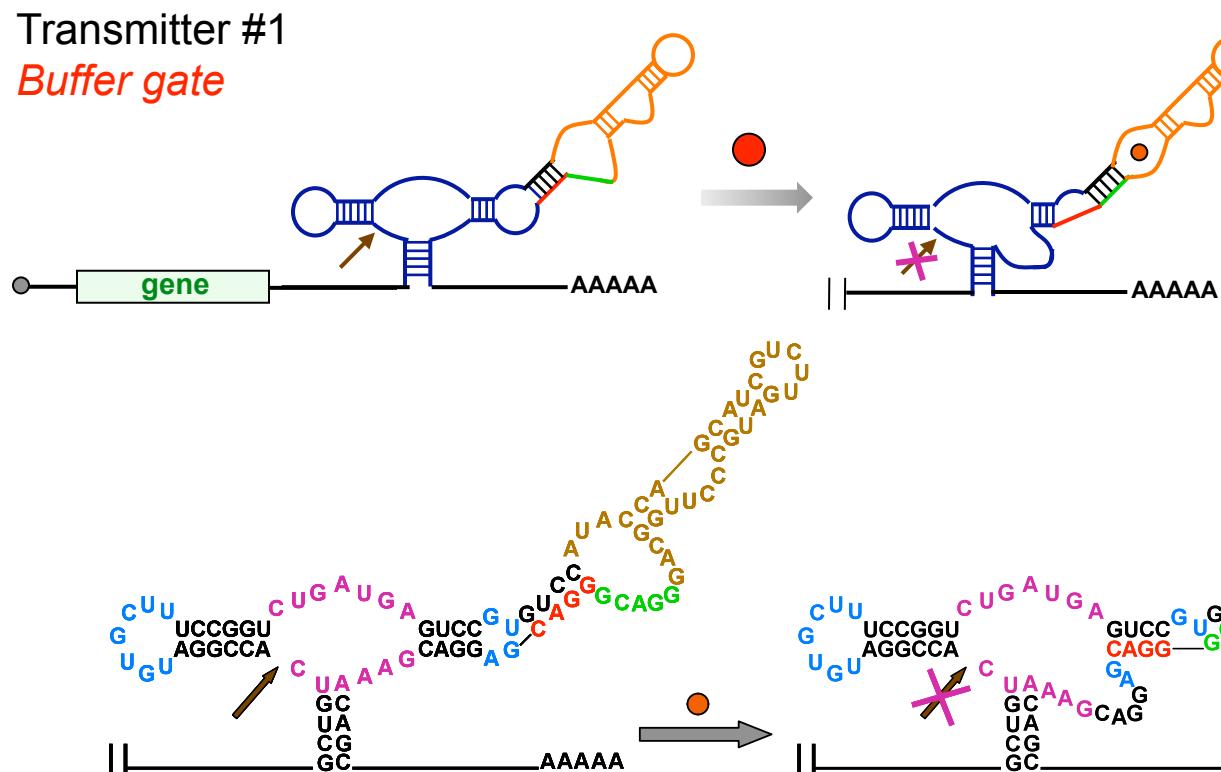
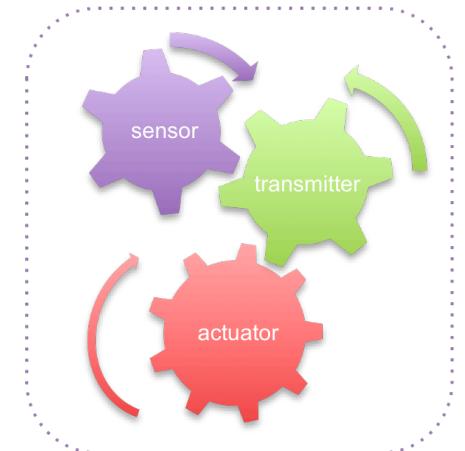
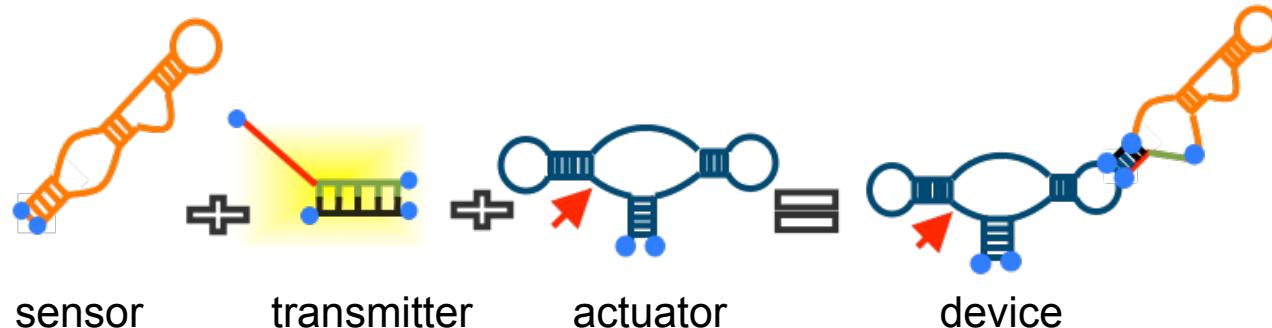


Transmitter #1

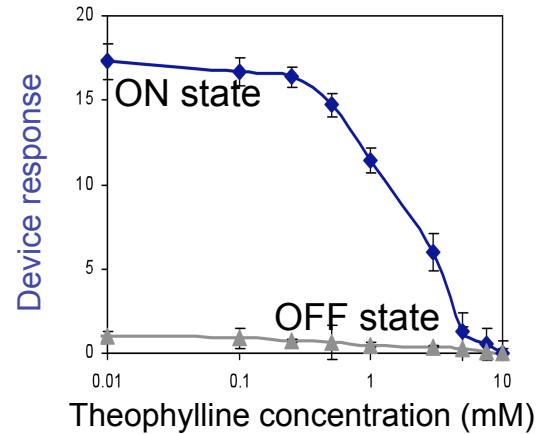
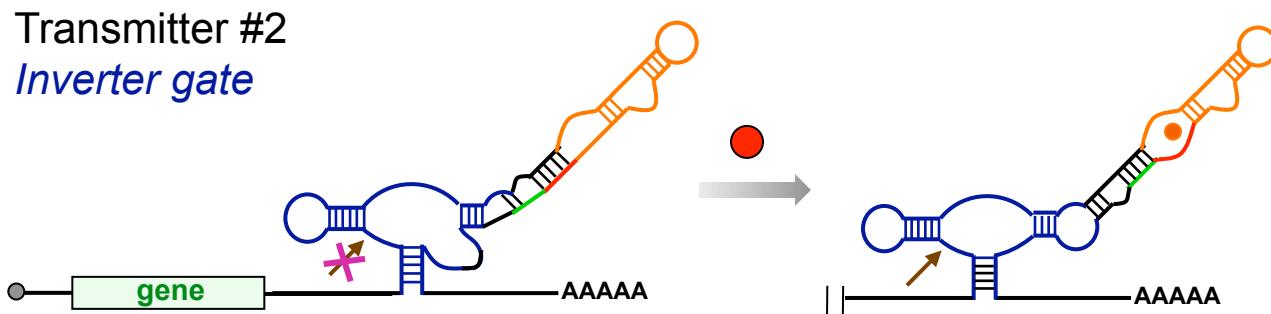
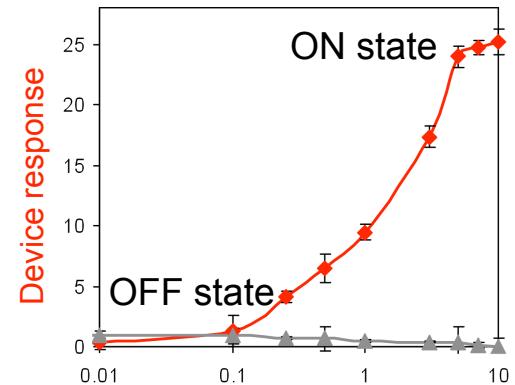
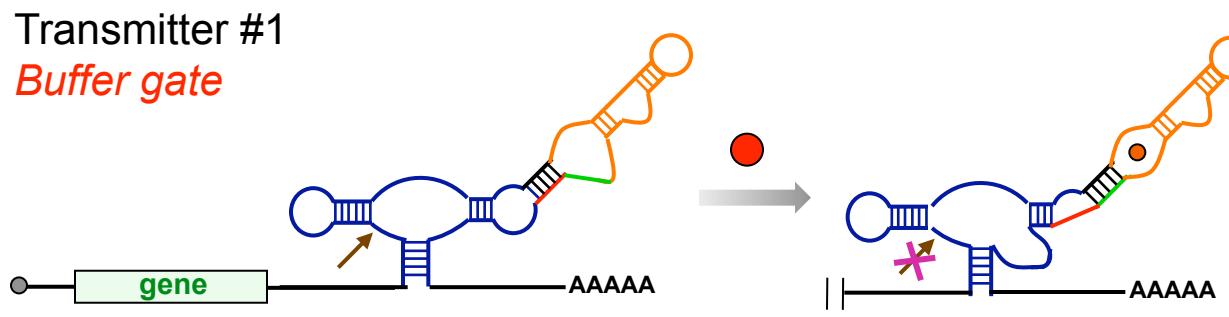
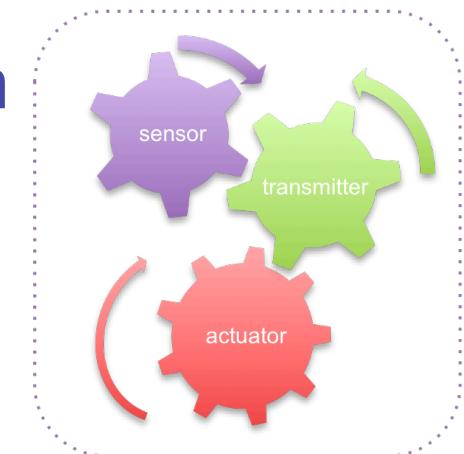
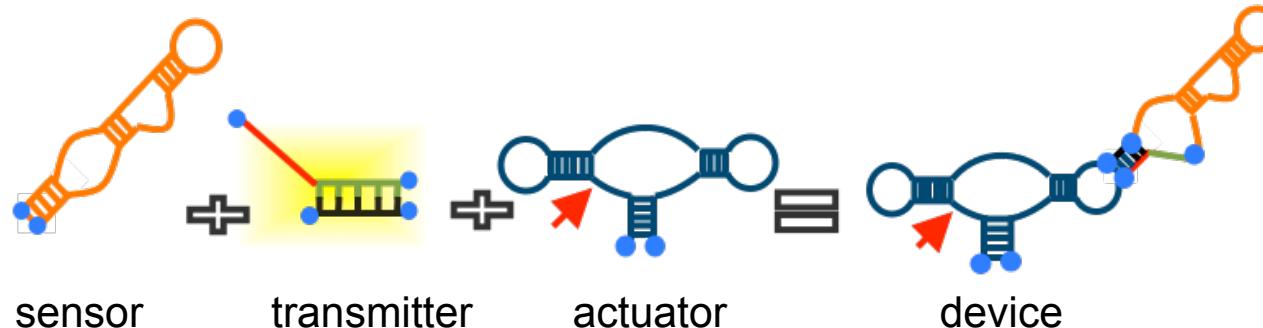
*Buffer gate*



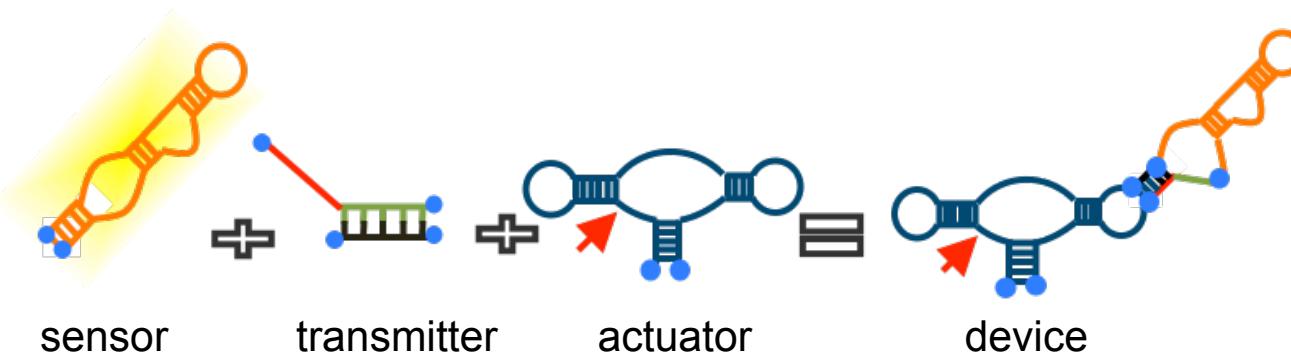
# A platform for programming device function



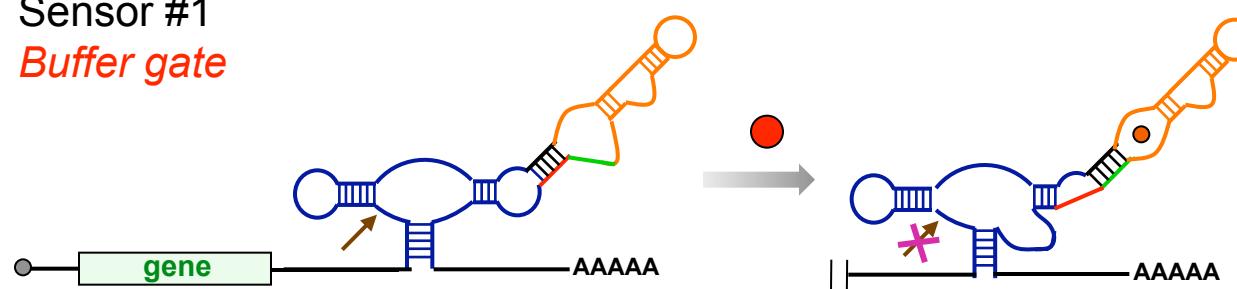
# A platform for programming device function



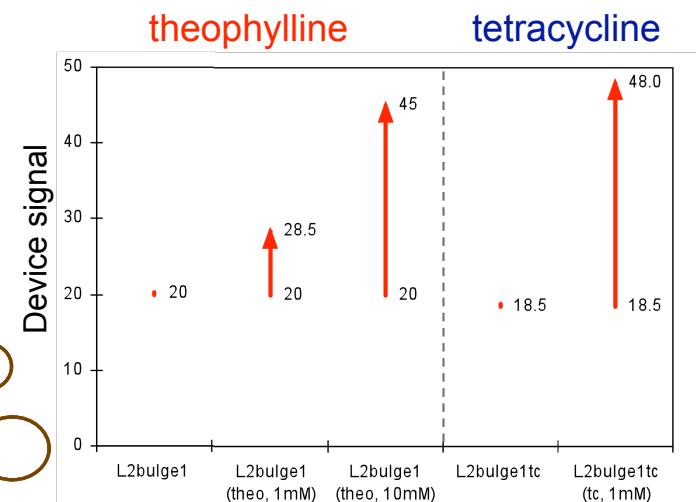
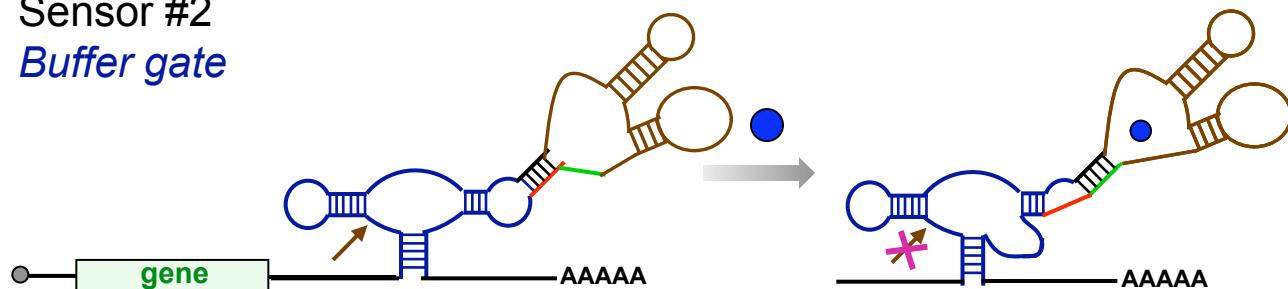
# A platform for programming device inputs



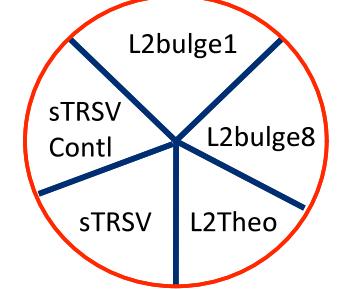
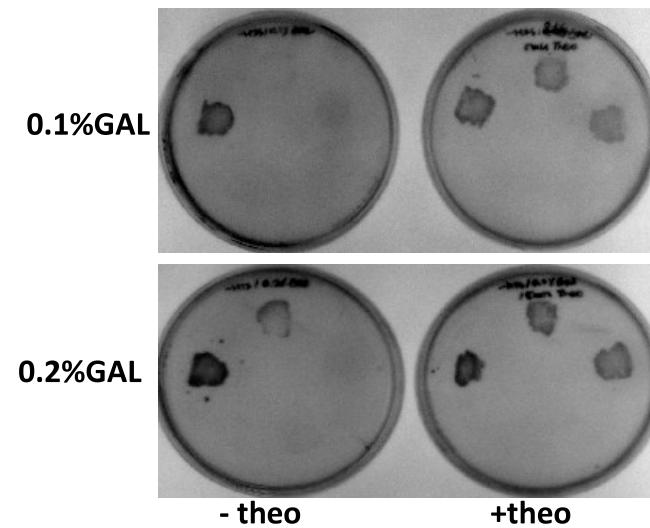
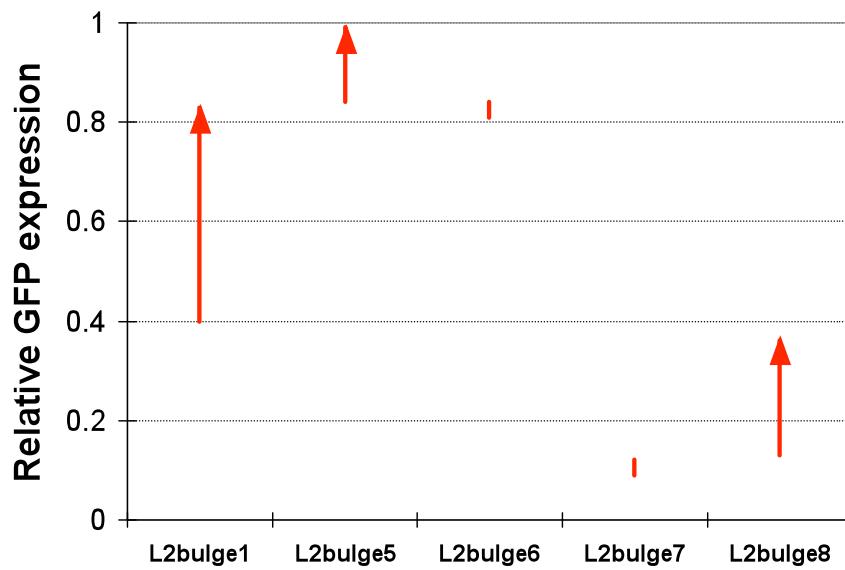
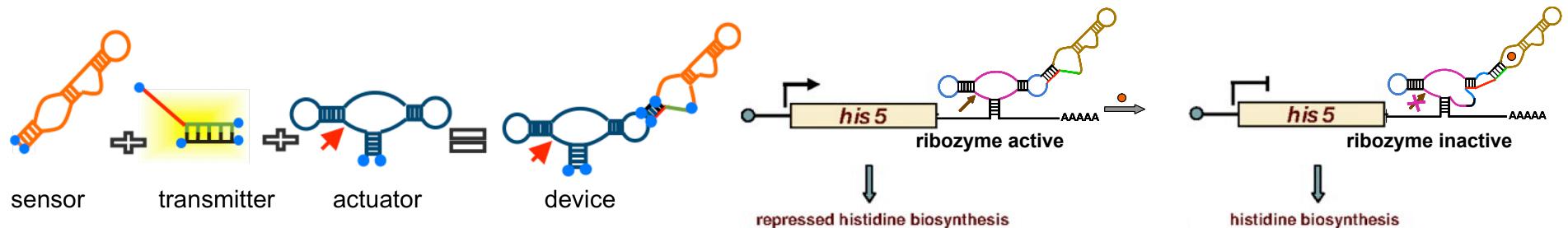
Sensor #1  
*Buffer gate*



Sensor #2  
*Buffer gate*



# A platform for programming response properties

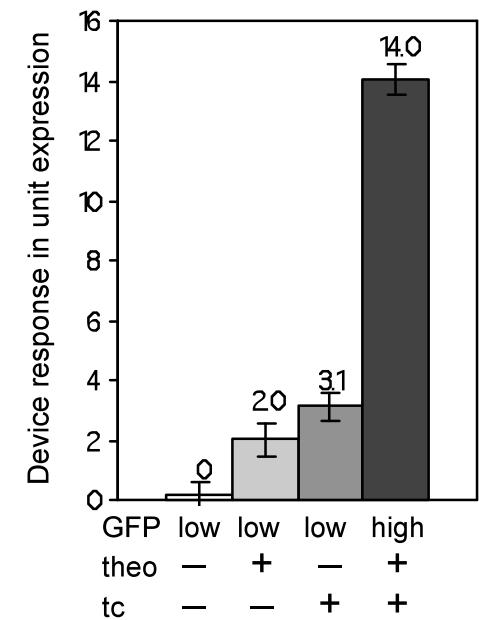
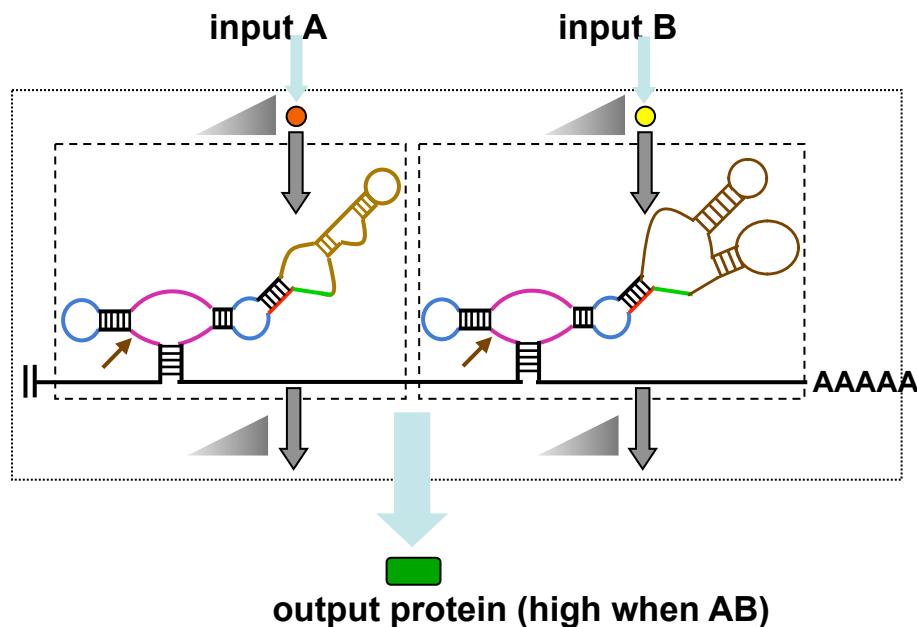


# Logic gate devices

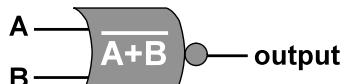
## AND gate



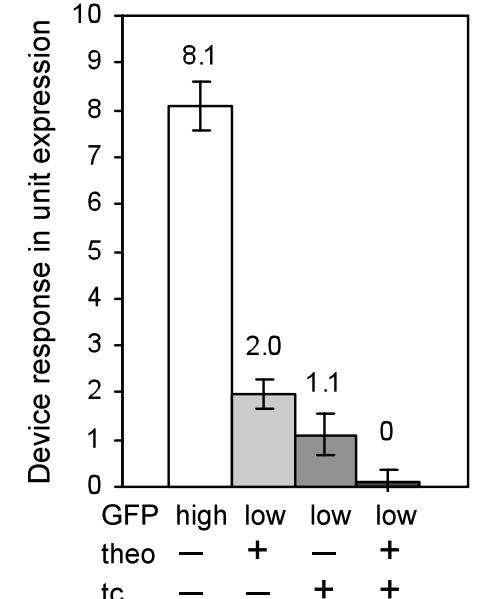
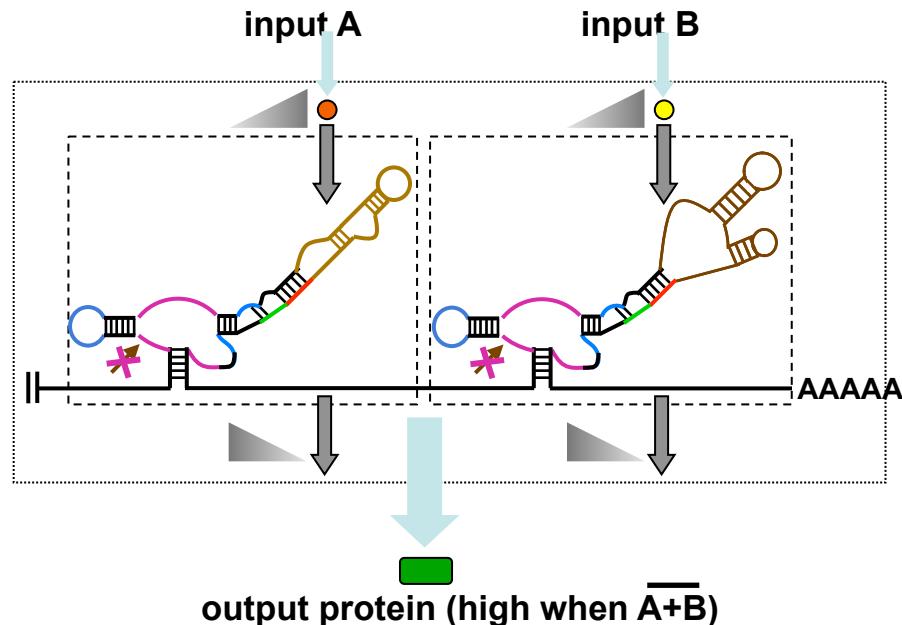
A	B	output
theo	tc	GFP
0	0	0
0	1	0
1	0	0
1	1	1



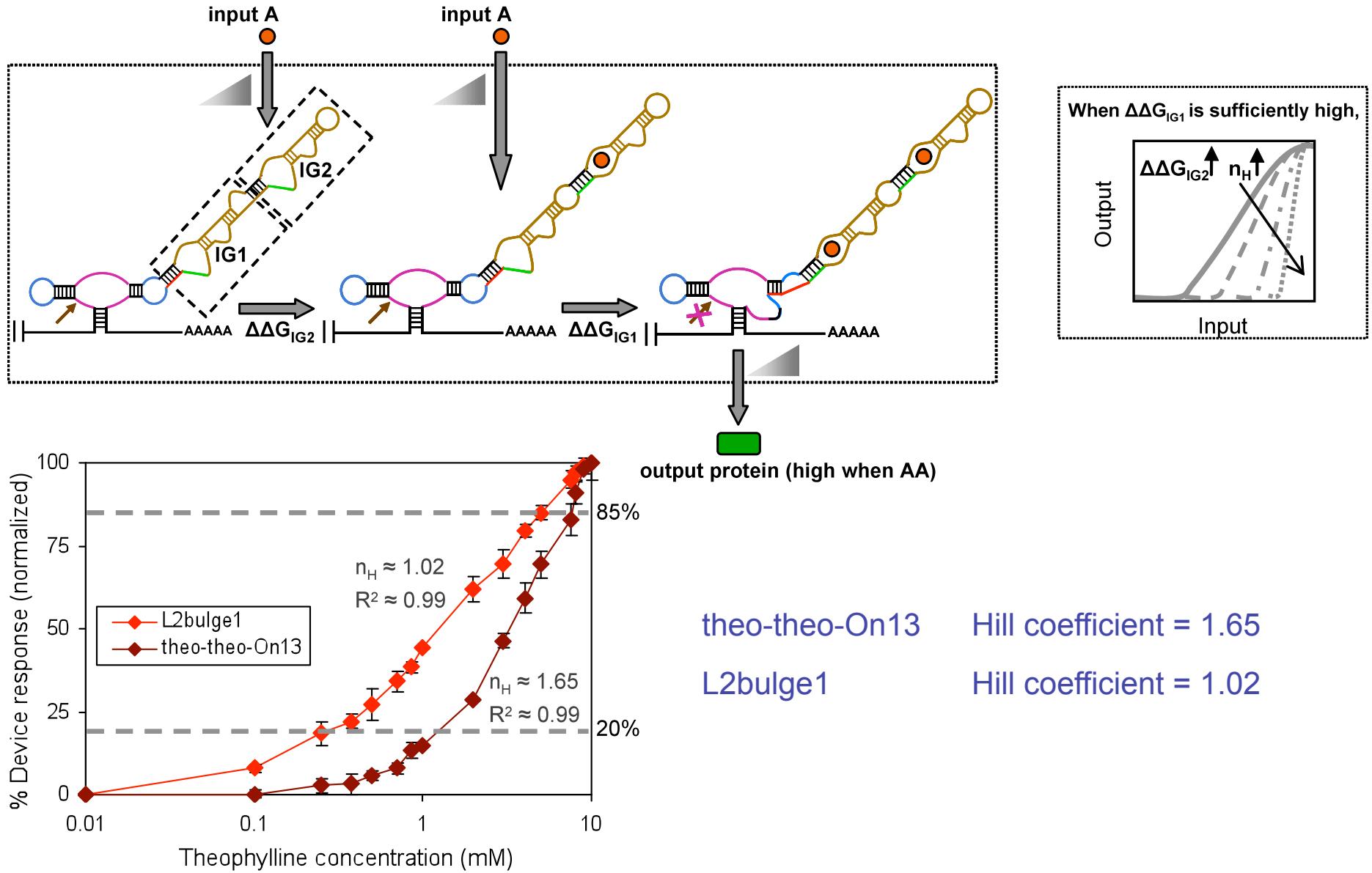
## NOR gate



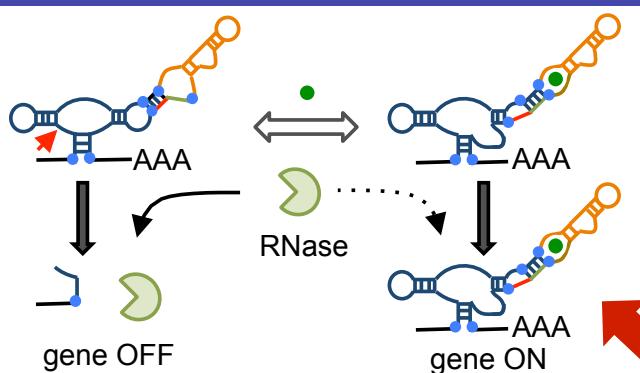
A	B	output
theo	tc	GFP
0	0	1
0	1	0
1	0	0
1	1	0



# Single gain device – programmed cooperativity

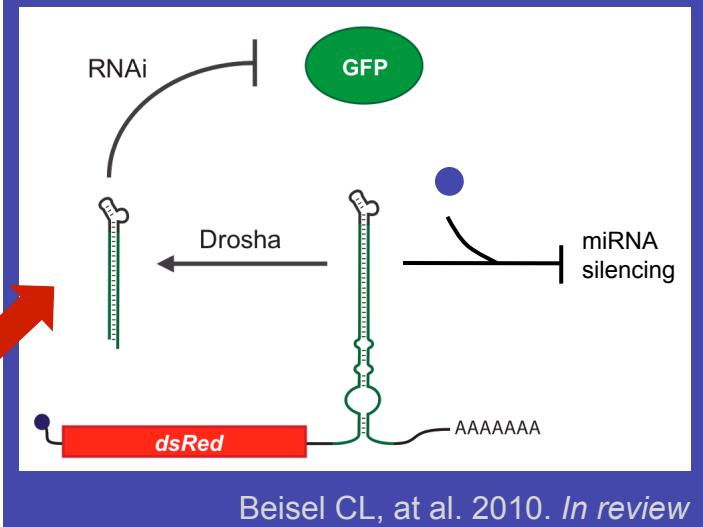


## Ribozyme Cleavage



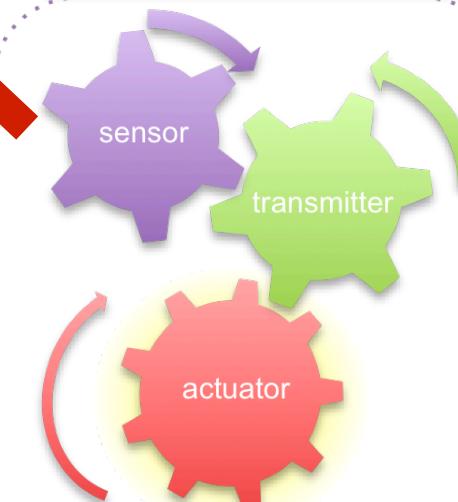
Win MN, Smolke CD. 2007. PNAS. 104: 14283-8  
Win MN, Smolke CD. 2008. Science. 322: 456-60

## miRNA-Mediated Silencing

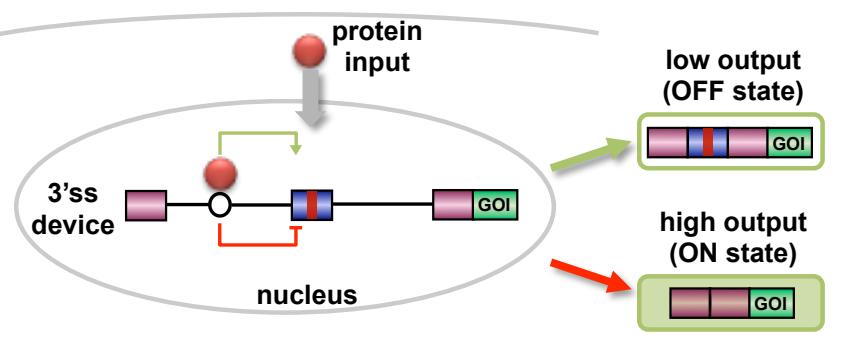


Beisel CL, et al. 2010. *In review*

## RNA devices

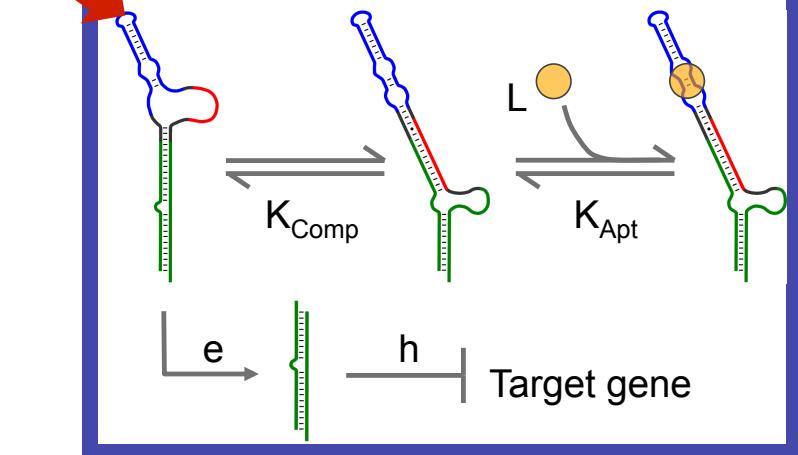


## Alternative Splicing



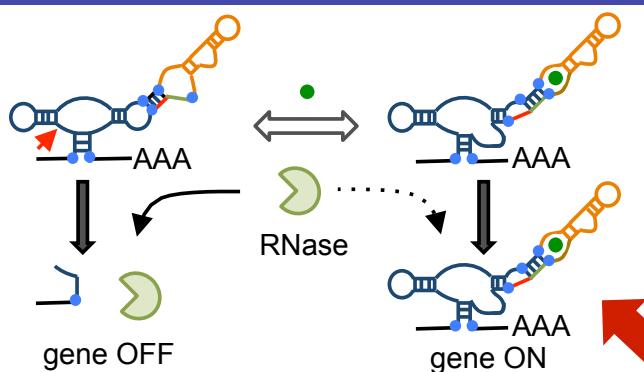
Culler SJ, Hoff KG, Smolke CD. 2010. *In review*

## shRNA-Mediated Silencing

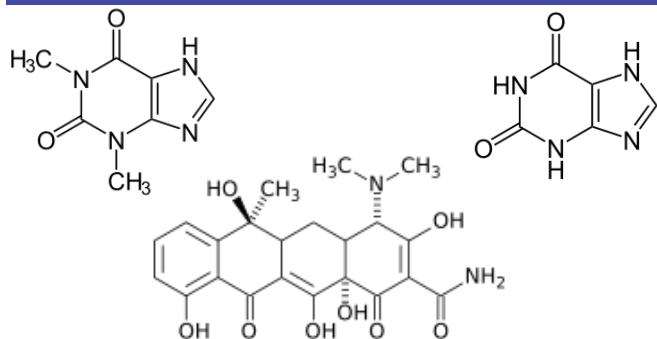


Beisel CL, et al. 2008. Mol Sys Biol. 4: 224

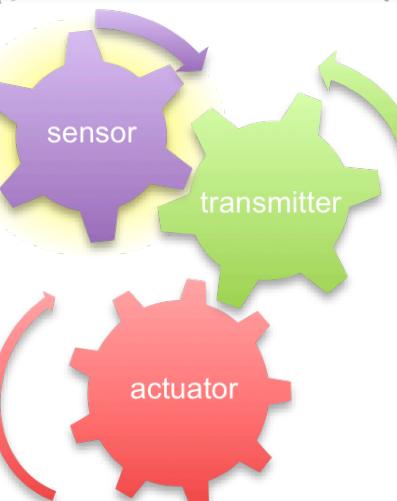
## Ribozyme Cleavage



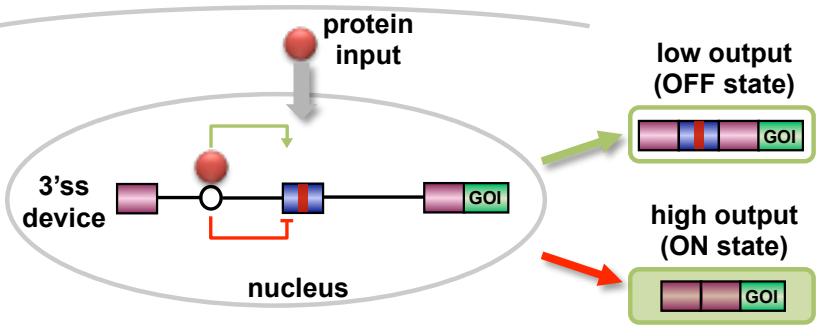
Win MN, Smolke CD. 2007. PNAS. 104: 14283-8  
Win MN, Smolke CD. 2008. Science. 322: 456-60



## RNA devices

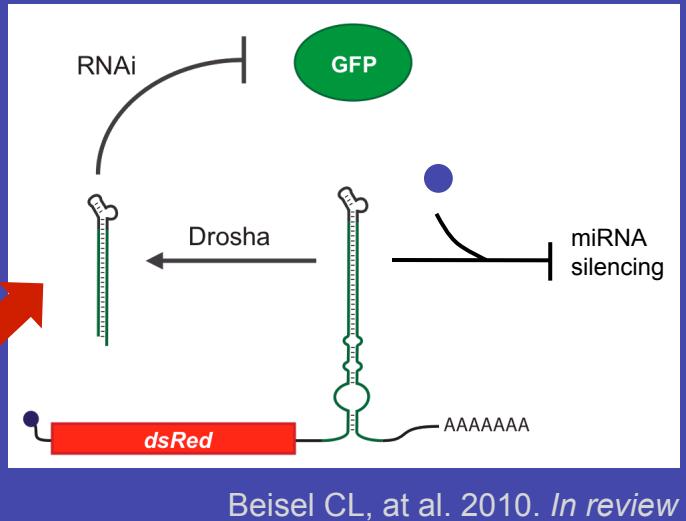


## Alternative Splicing



Culler SJ, Hoff KG, Smolke CD. 2010. In review

## miRNA-Mediated Silencing



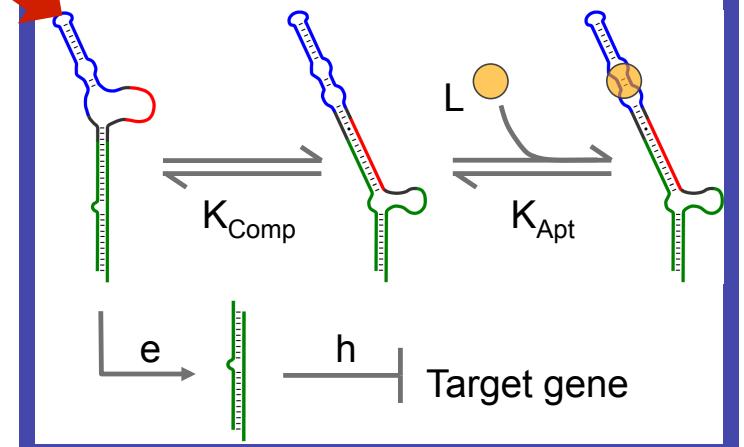
p50

$\beta$ -cat

p65

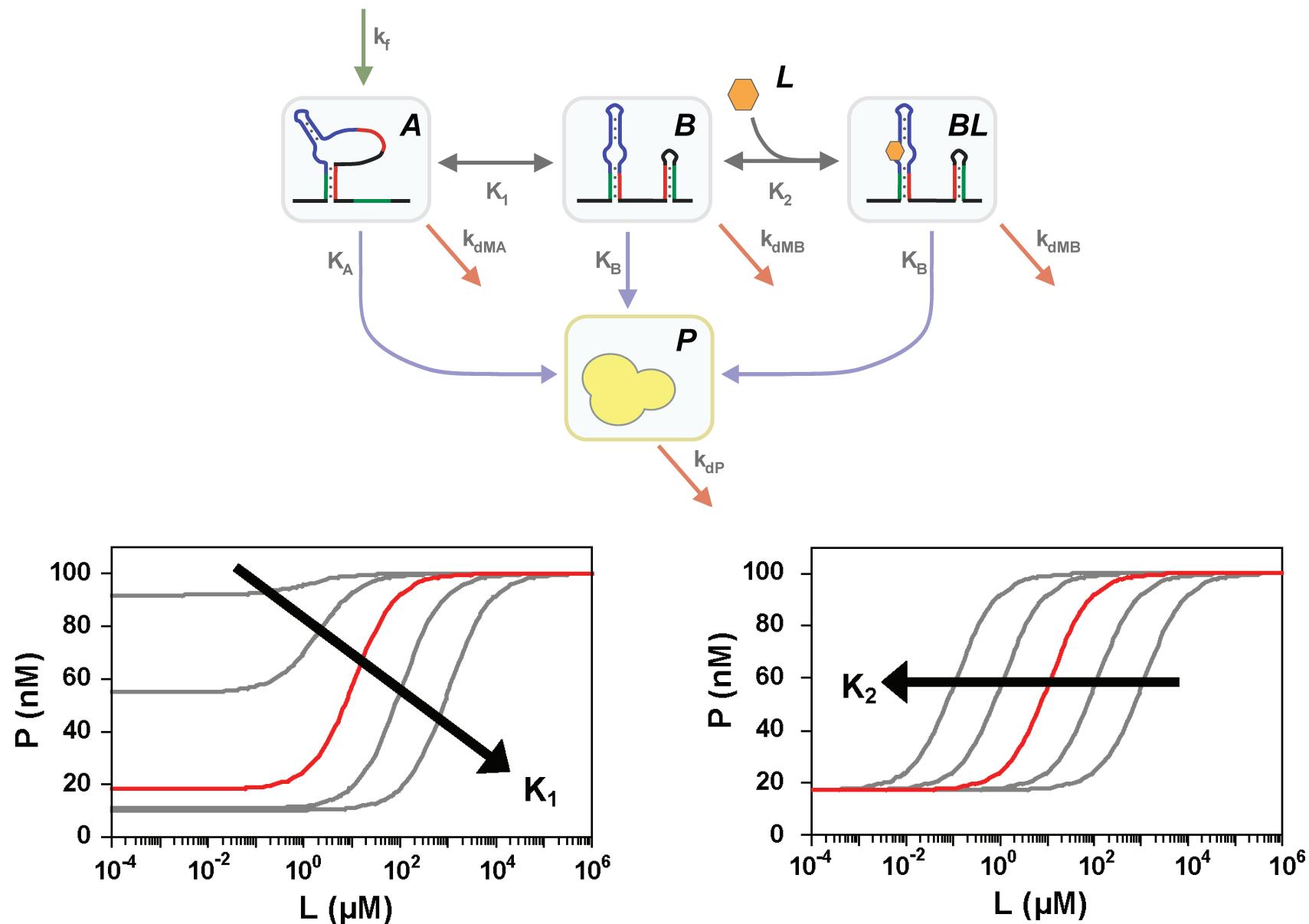
MS2

## shRNA-Mediated Silencing

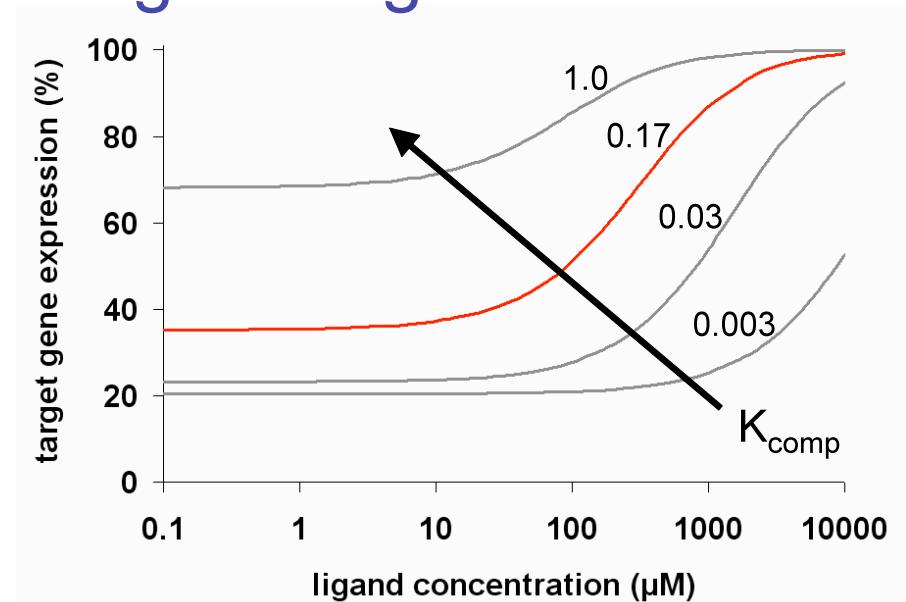
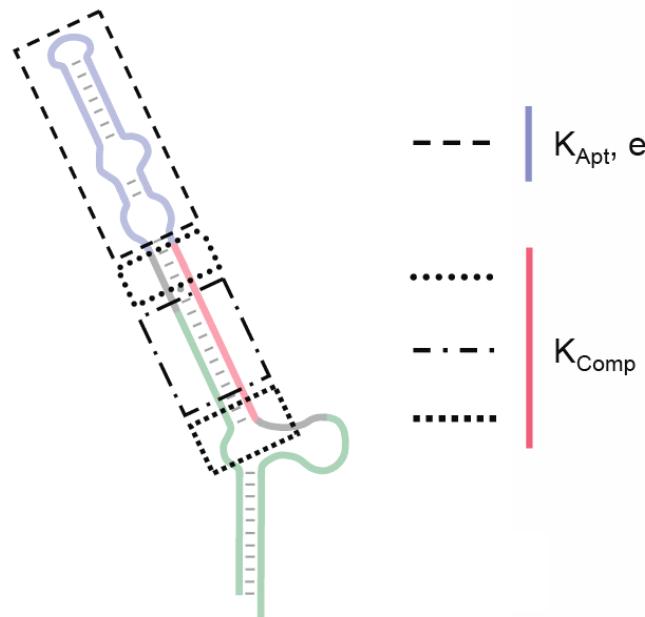


Beisel CL, et al. 2008. Mol Sys Biol. 4: 224

# Computational models can predict parameter effects

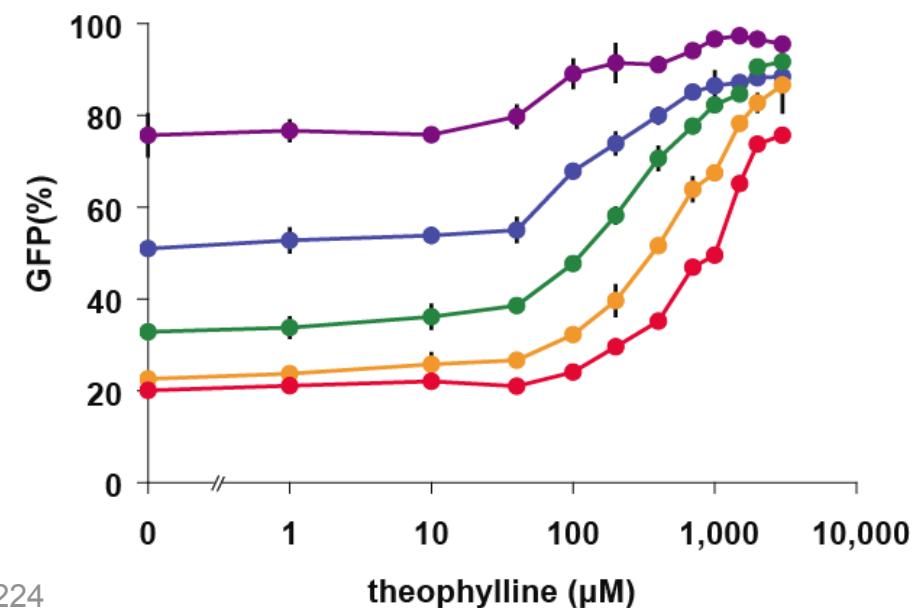
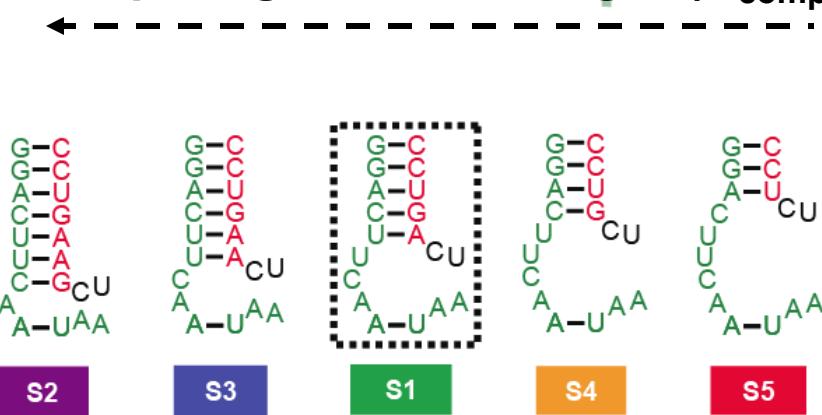


# Models predict transmitter tuning strategies

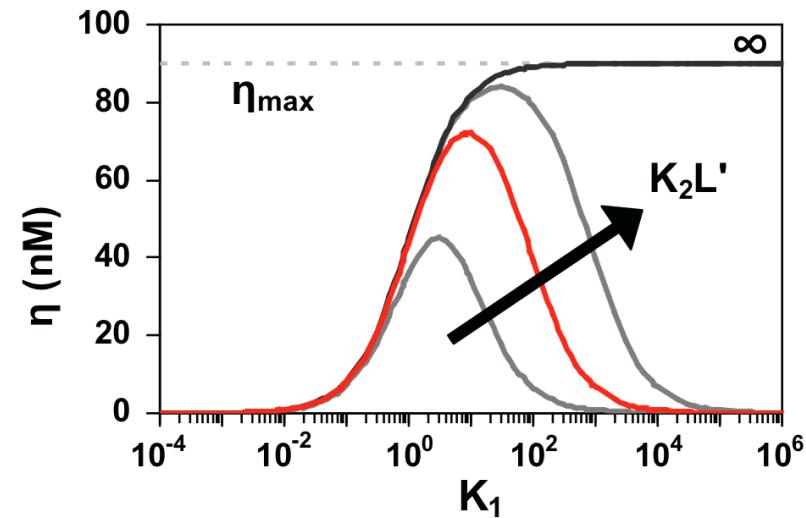
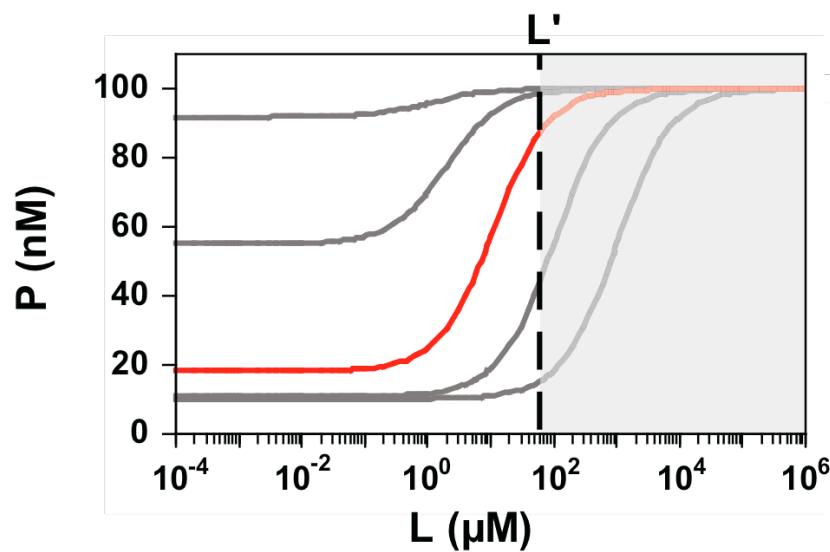
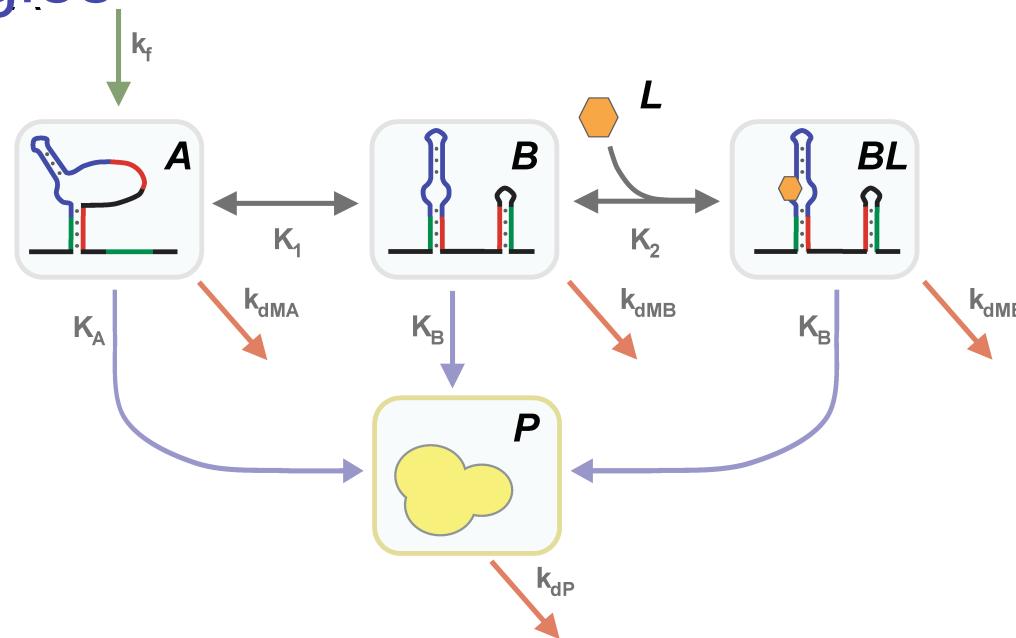


## Construction and experimental validation

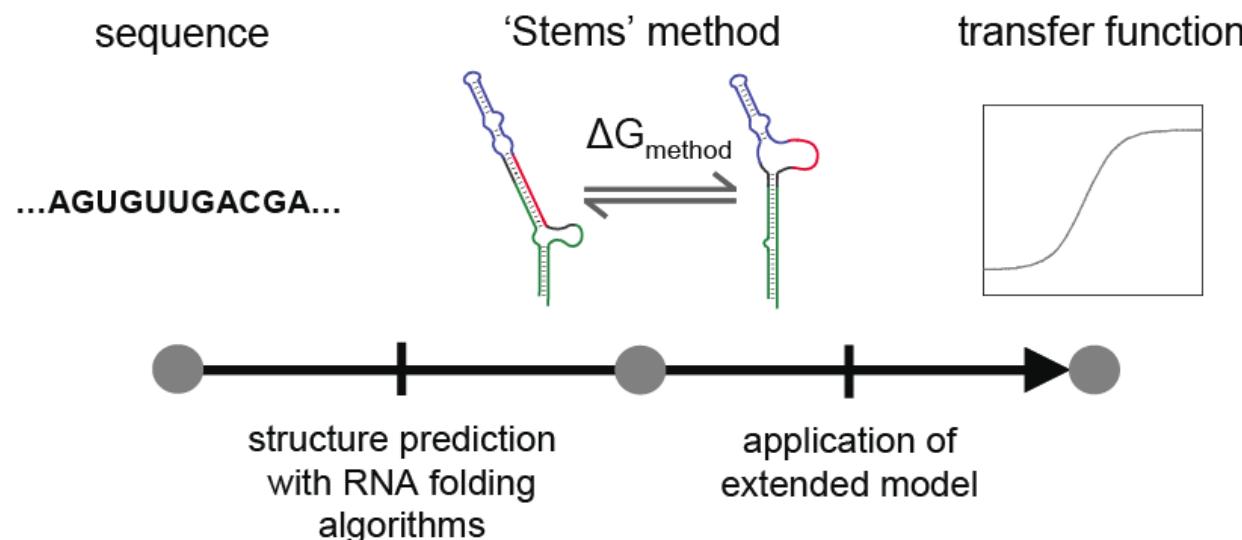
### Competing strand 3' length ( $K_{\text{comp}}$ )



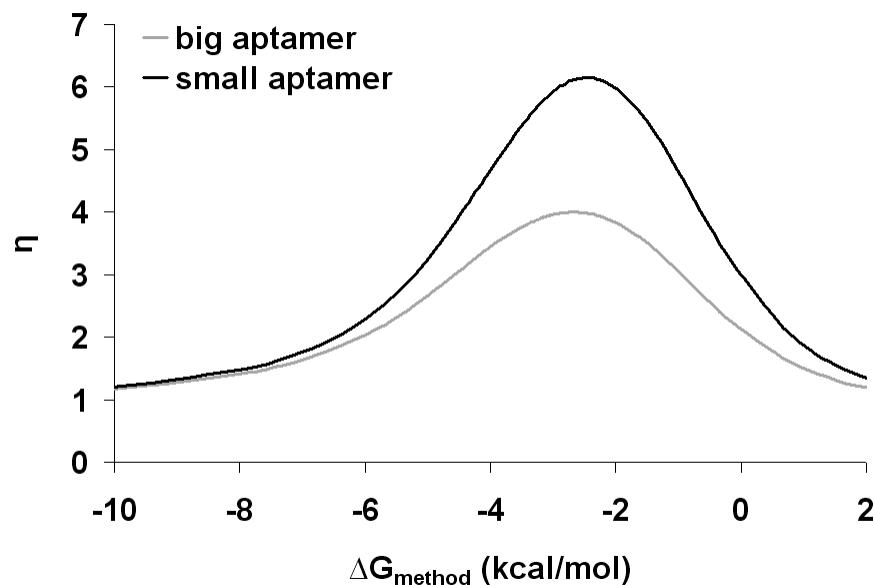
# System restrictions lead to optimization in RNA device design strategies



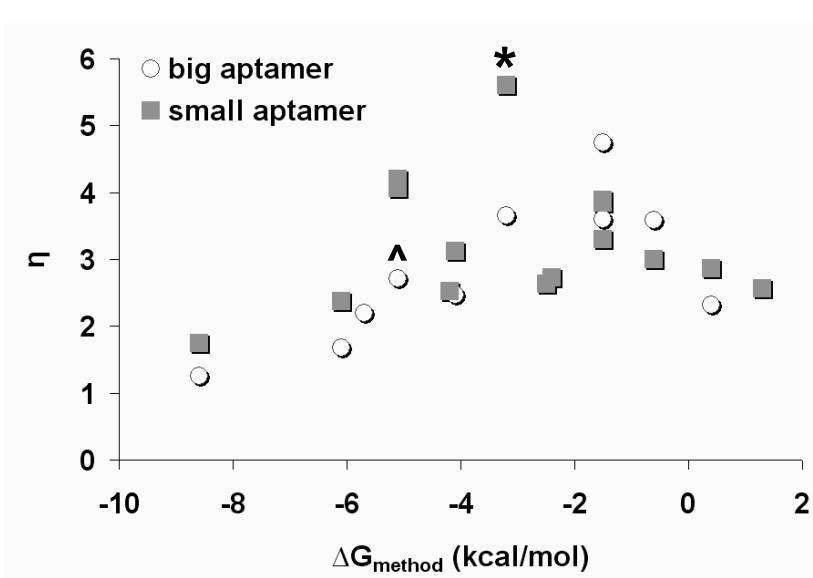
# A framework for forward engineering device response



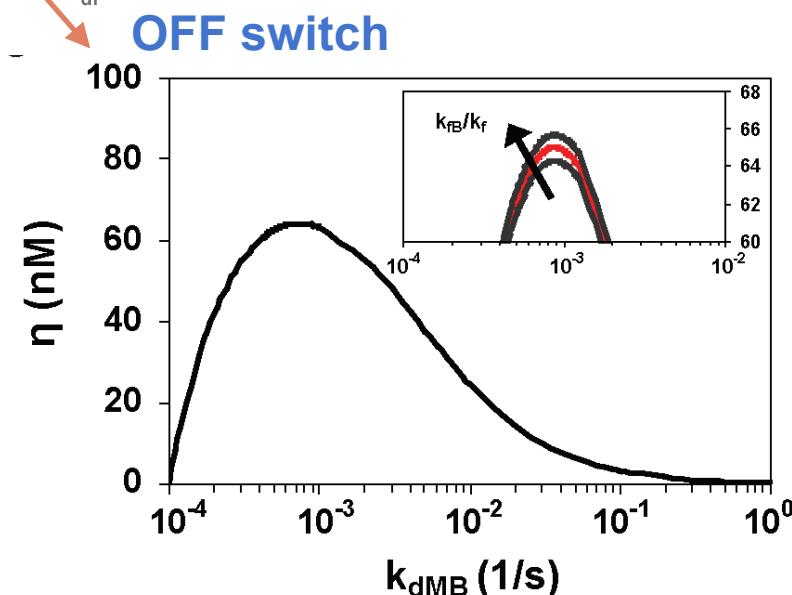
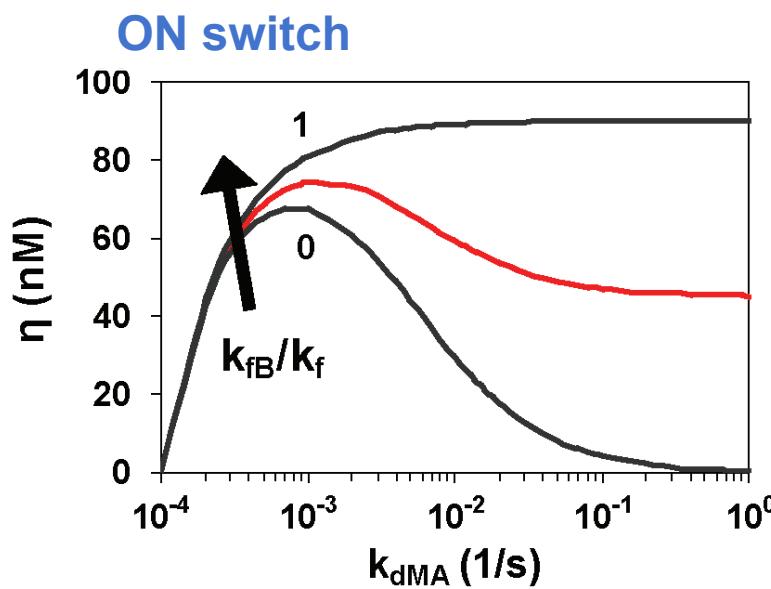
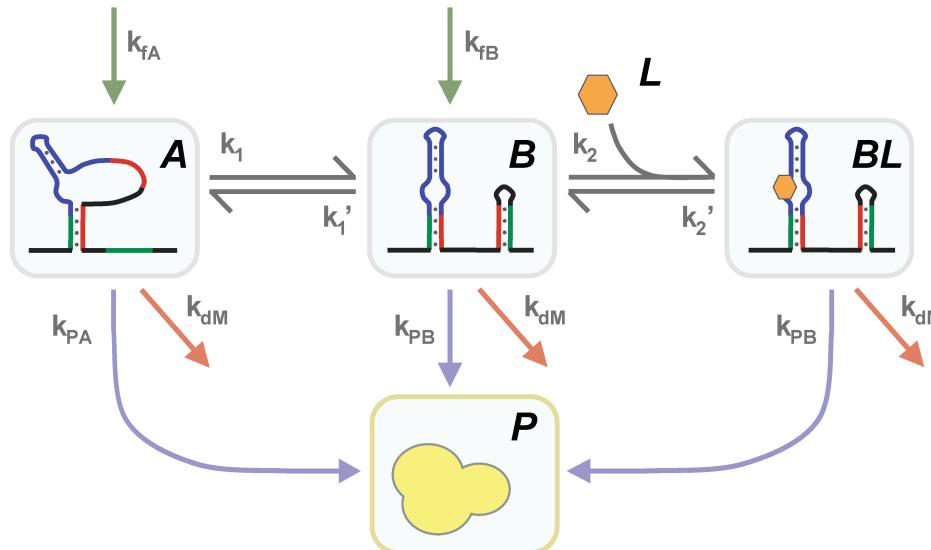
## Model predictions



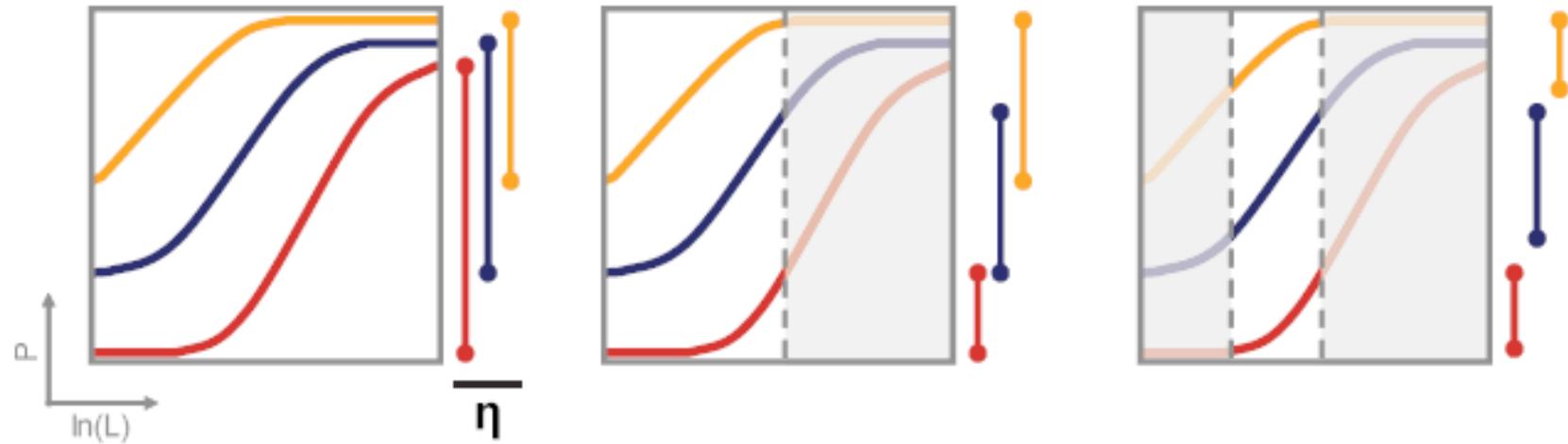
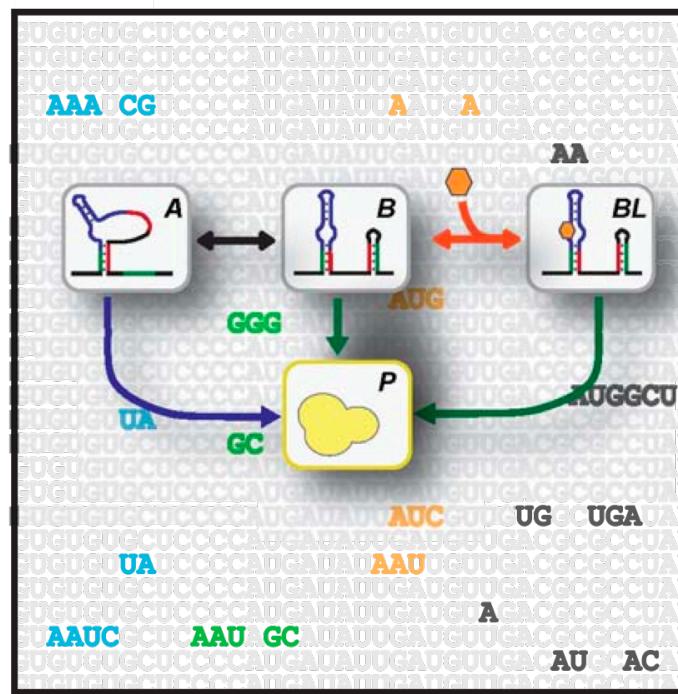
## Experimental results



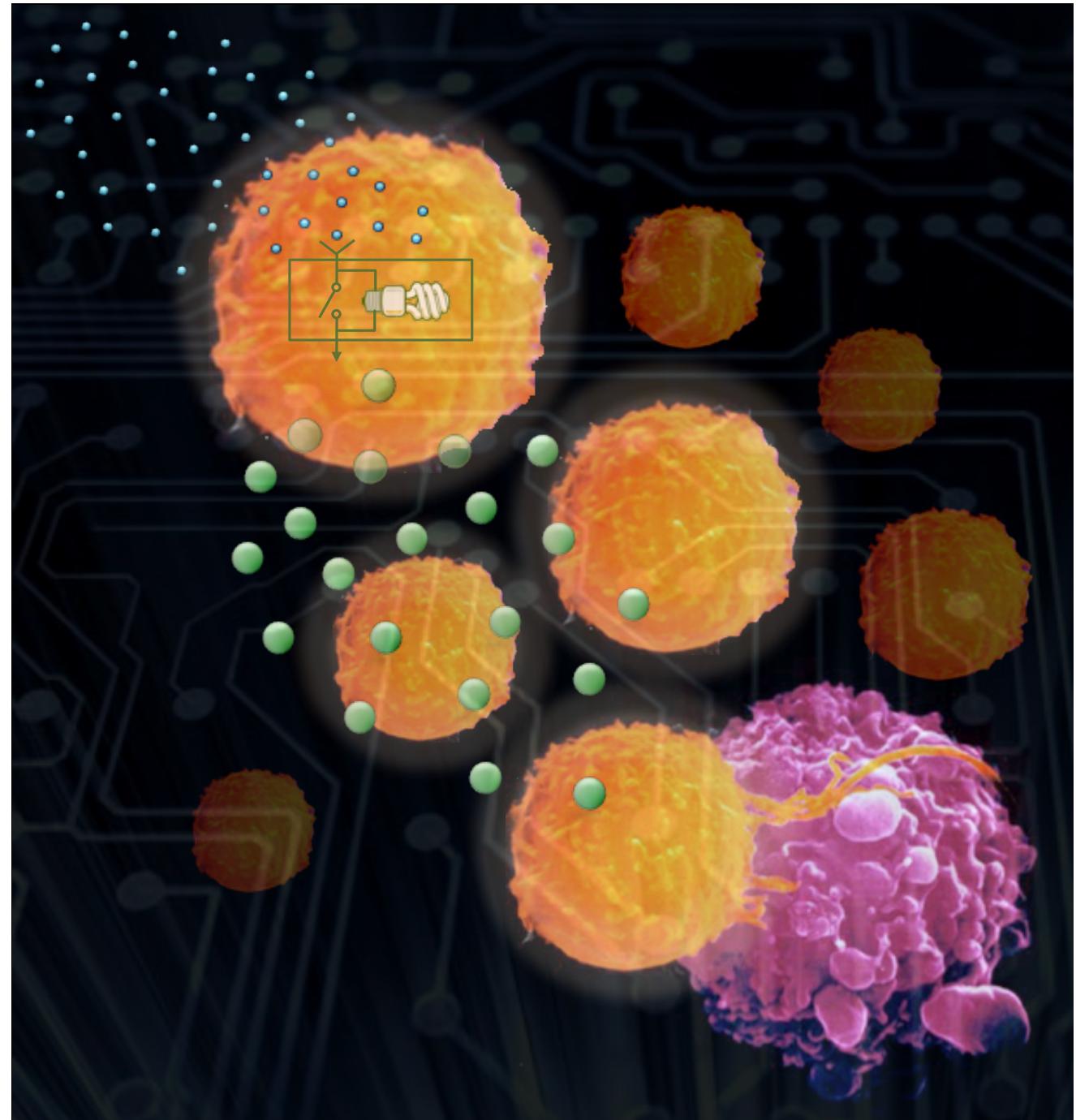
# Irreversible rates can play an important role in RNA device performance



# Quantitative tuning of device response

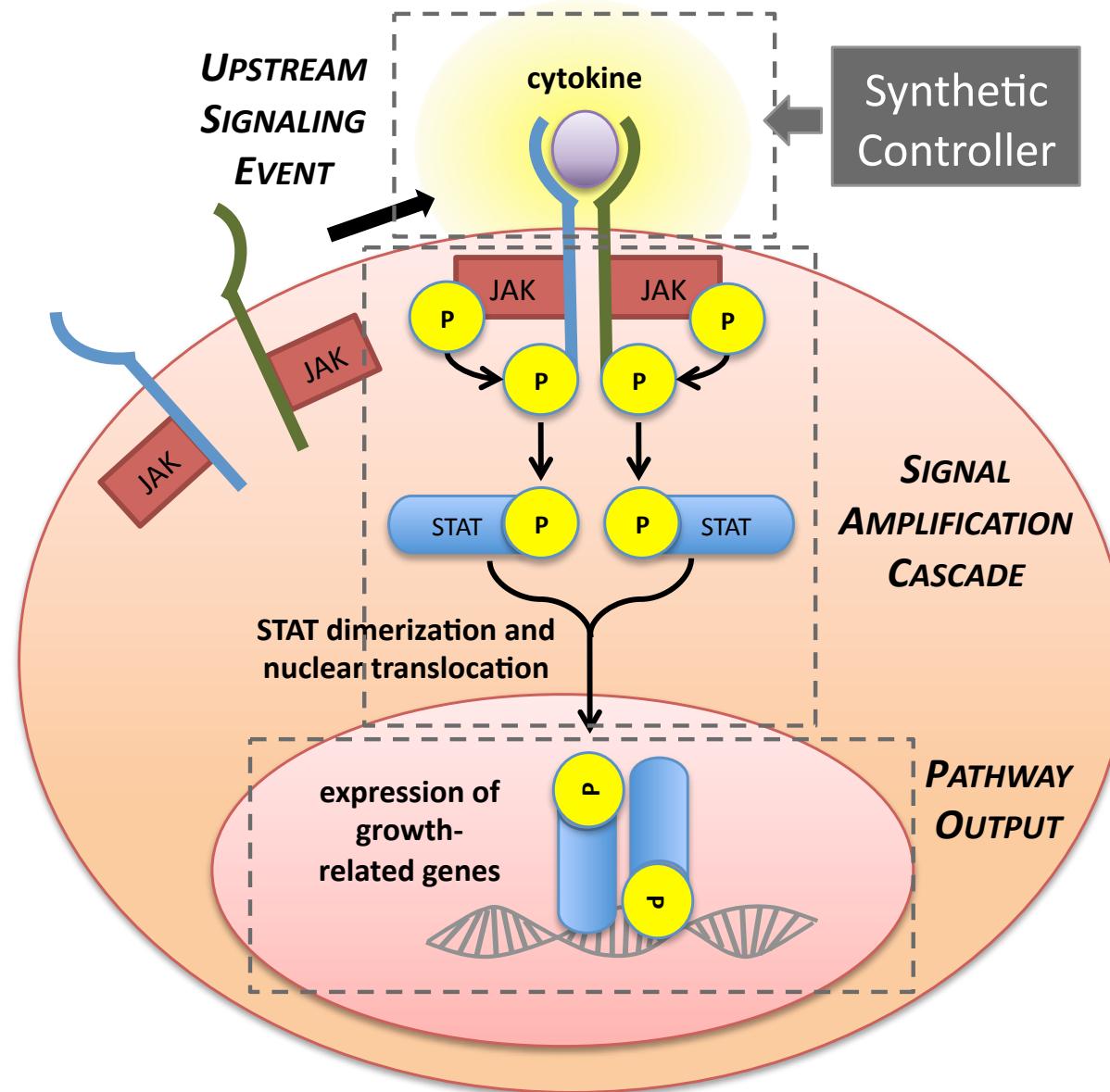


# Advancing cell-based therapies

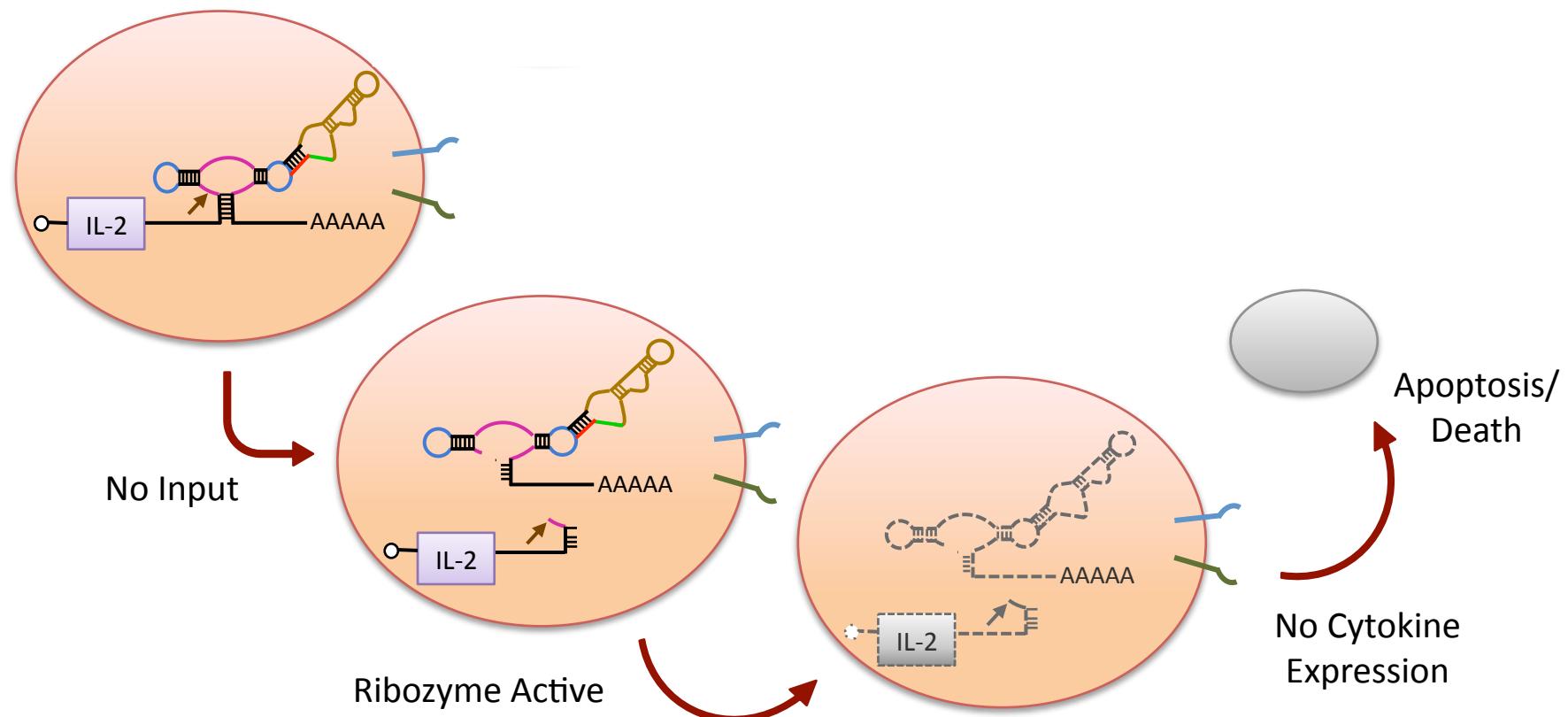


Chen YY, Jensen MC, Smolke CD. 2010. PNAS. 107: 8531-6

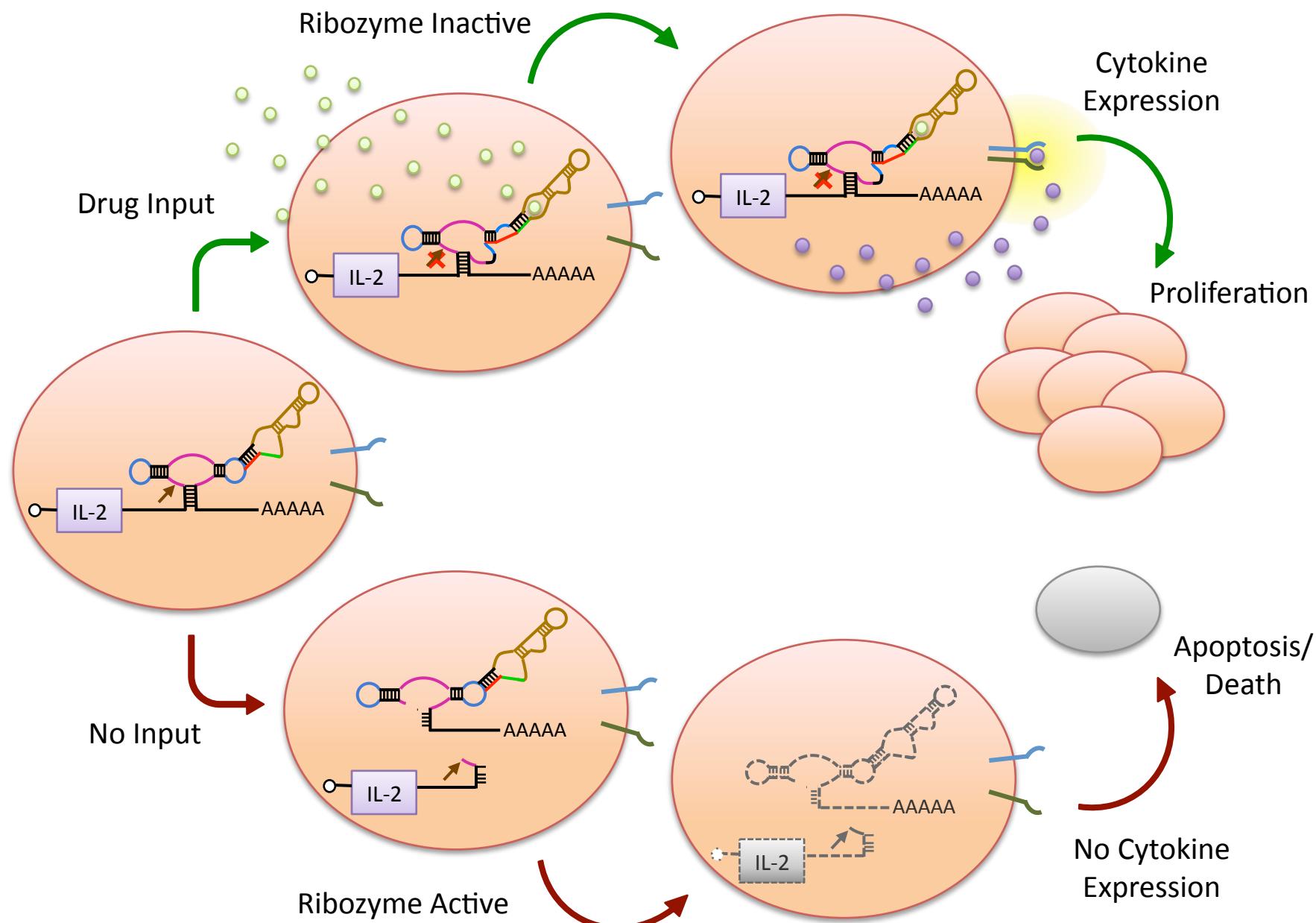
# T-cell proliferation pathway



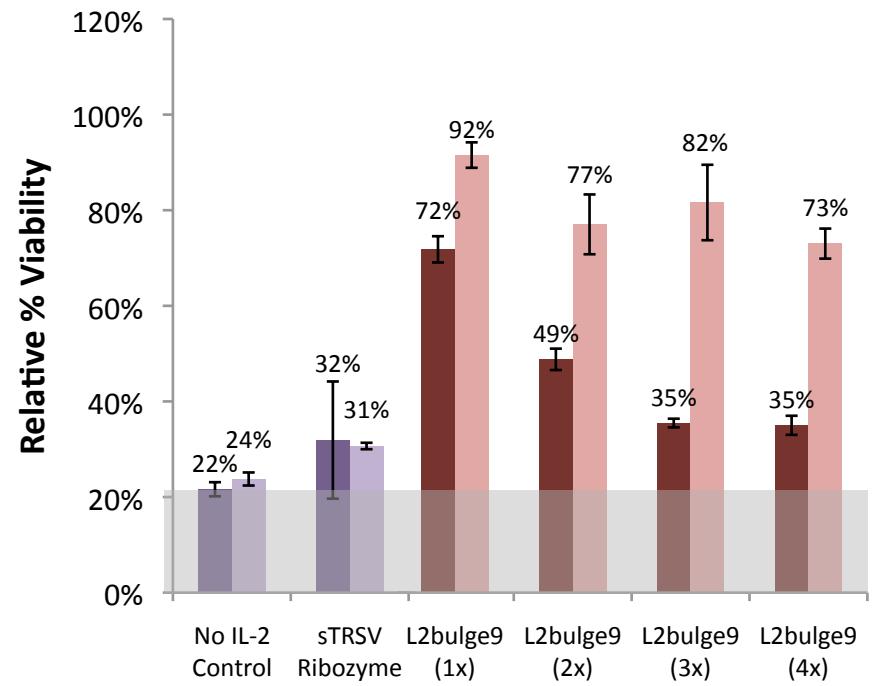
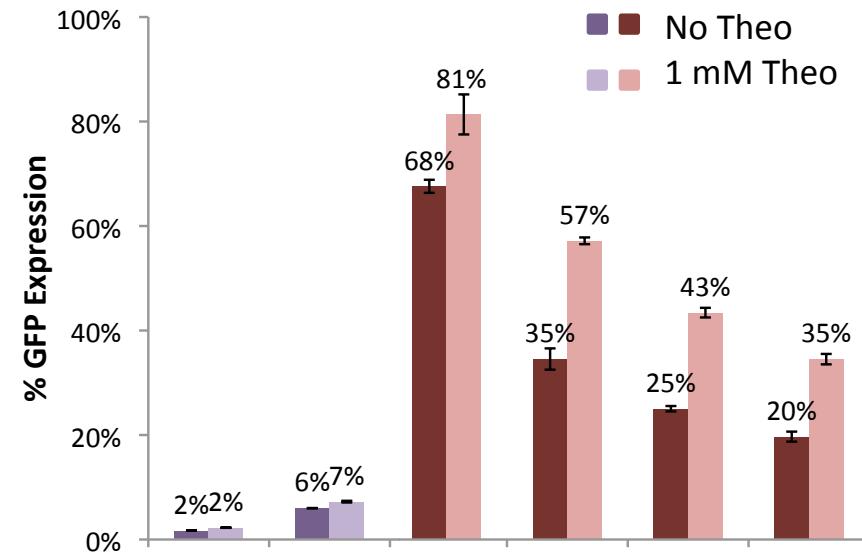
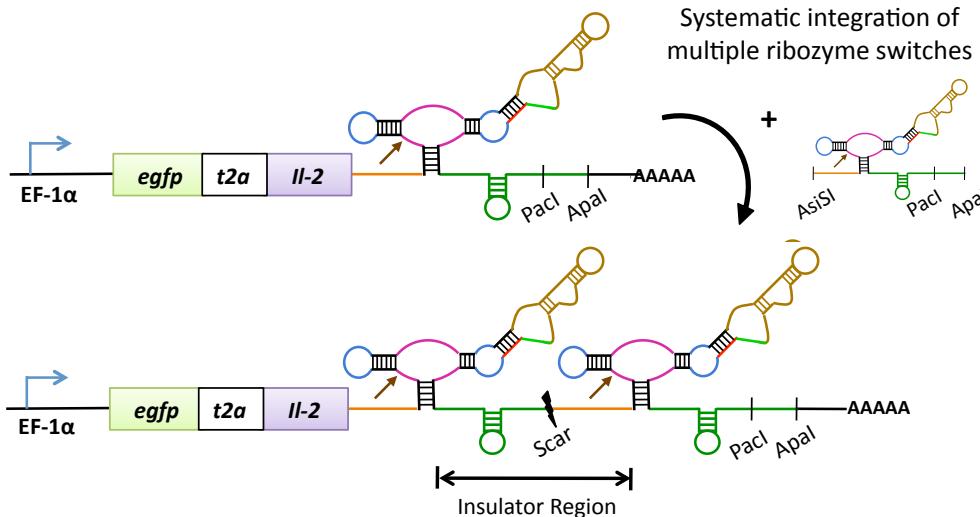
# A synthetic T-cell regulatory network



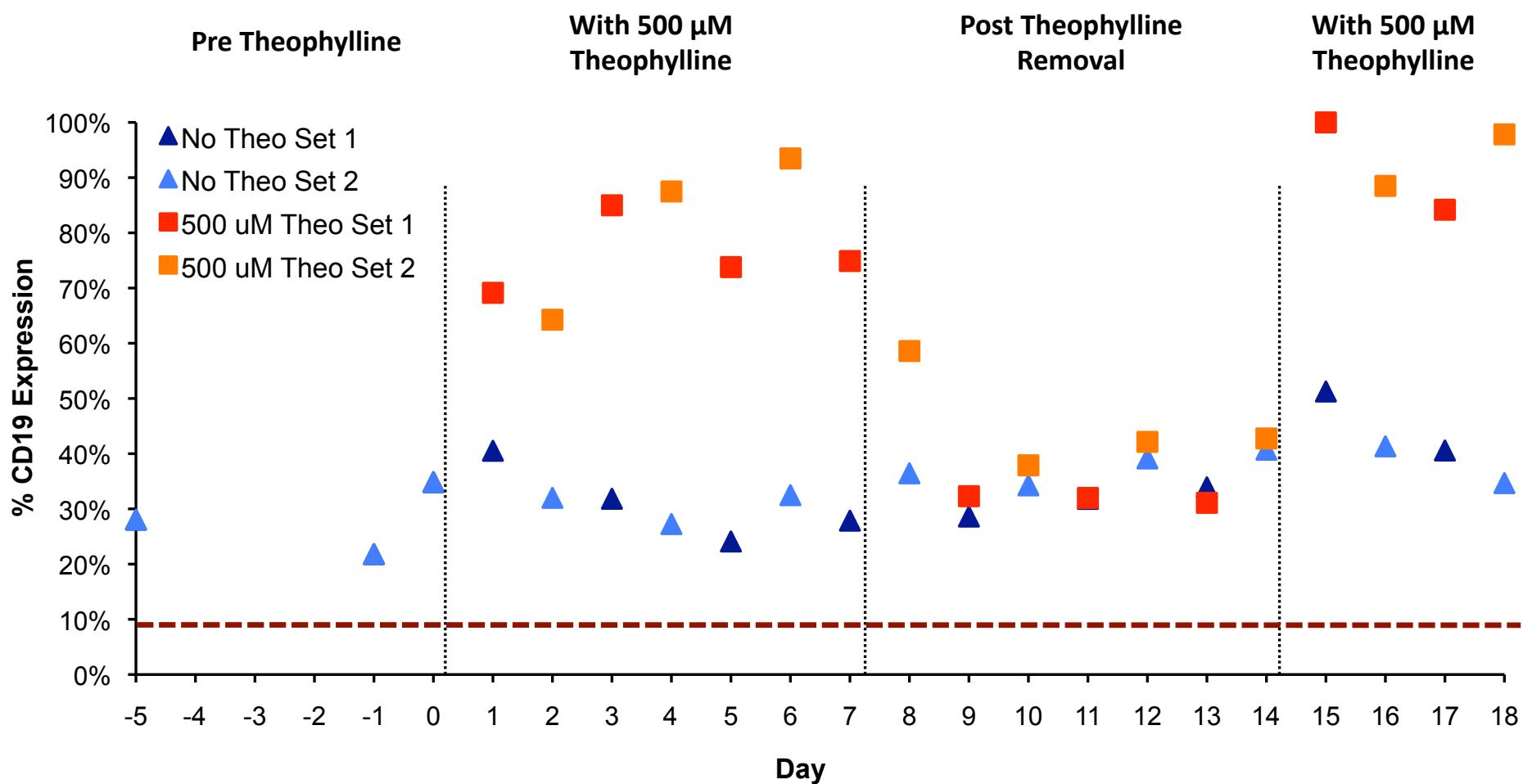
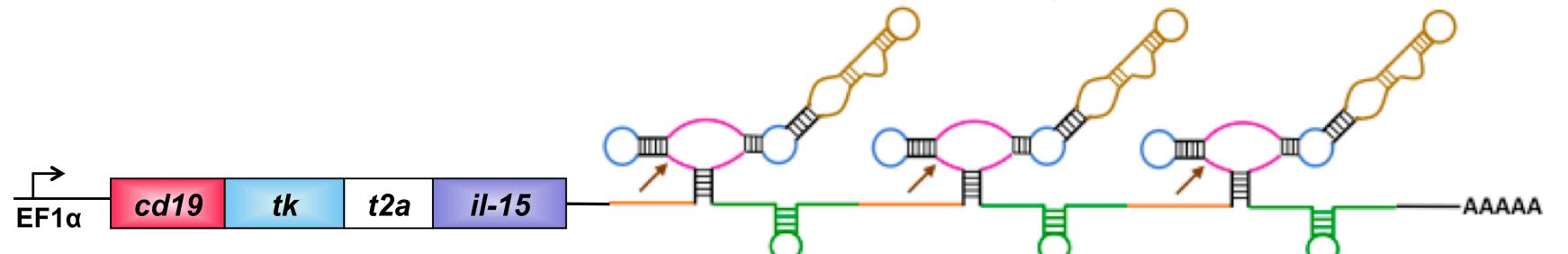
# A synthetic T-cell regulatory network



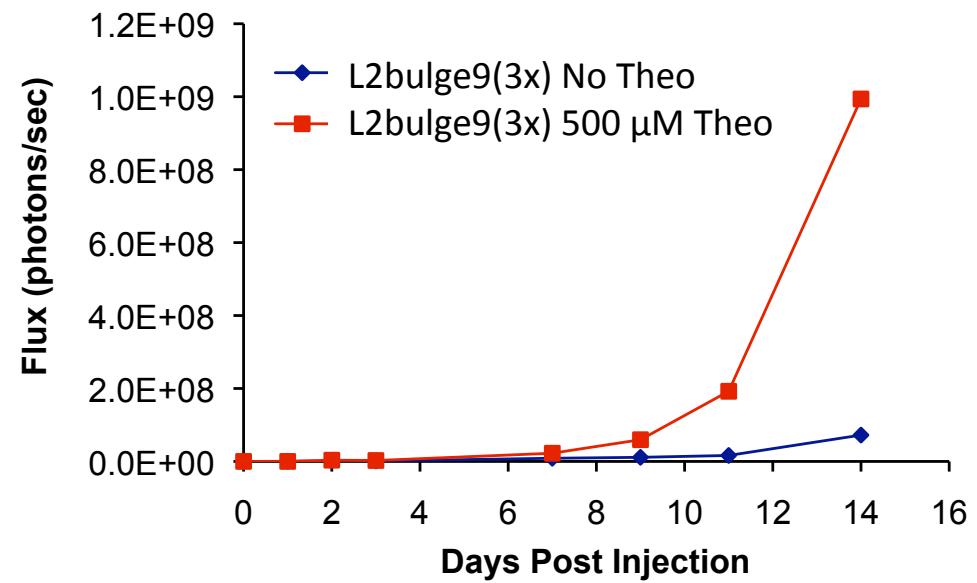
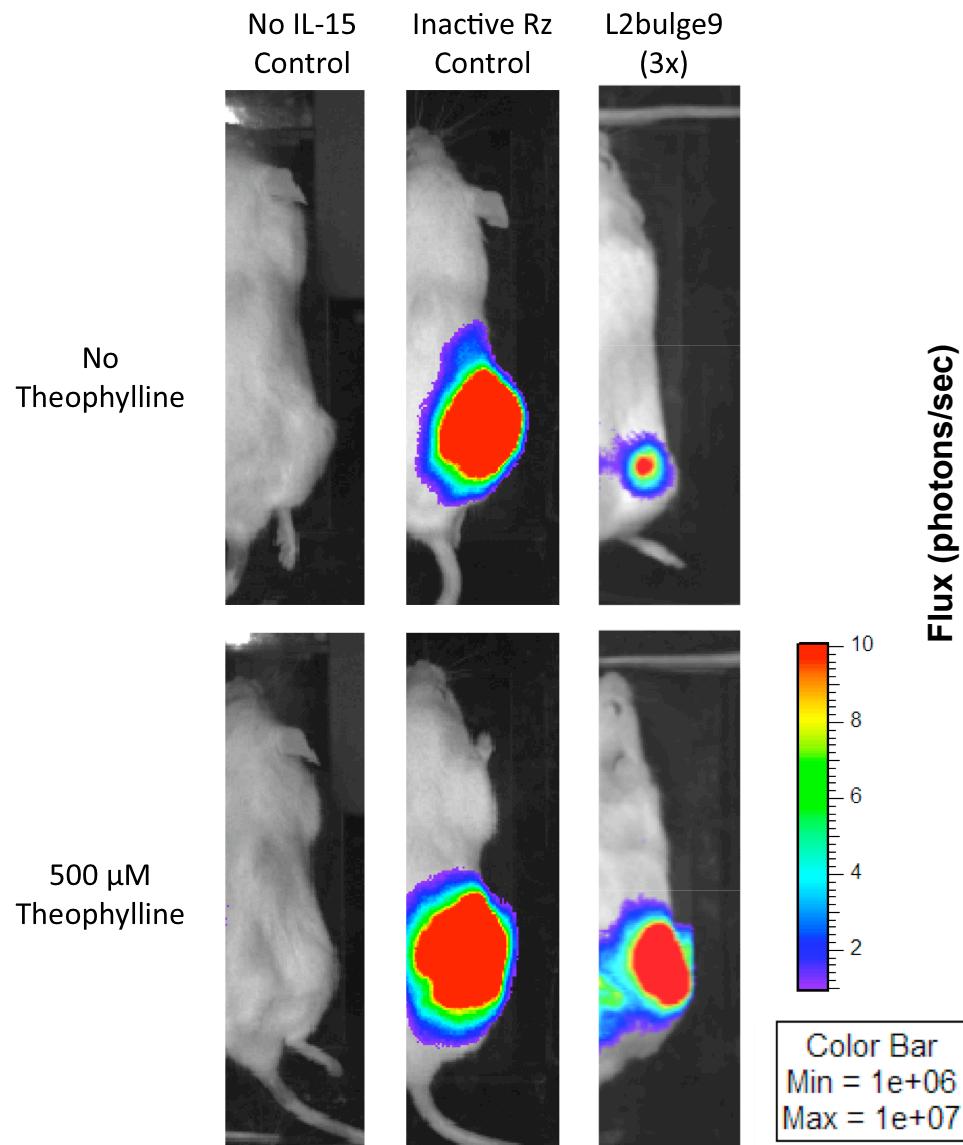
# Rapid prototyping and optimization of a T-cell proliferation control system



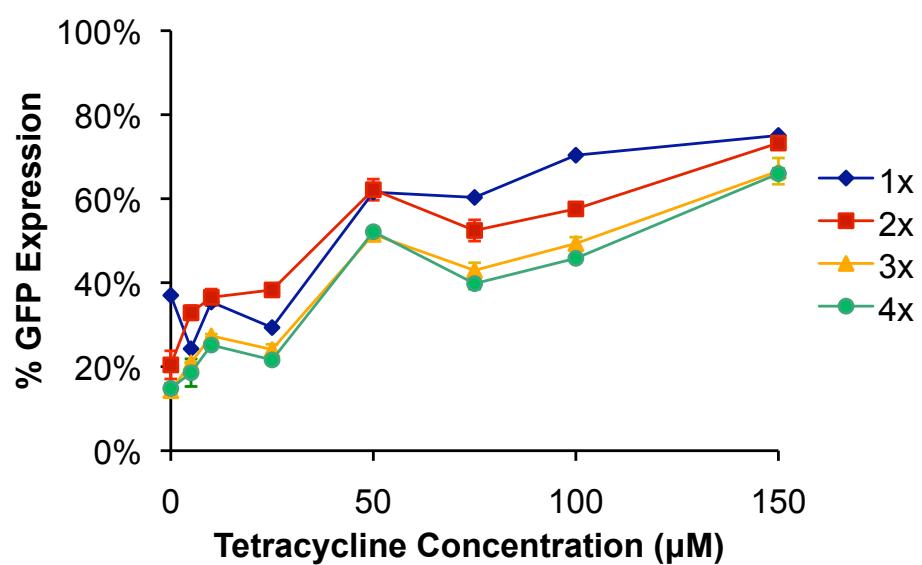
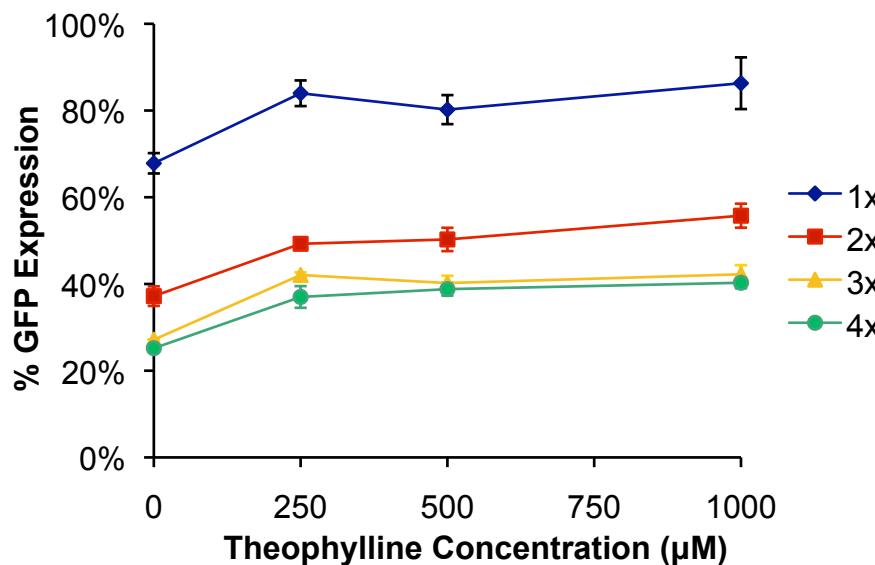
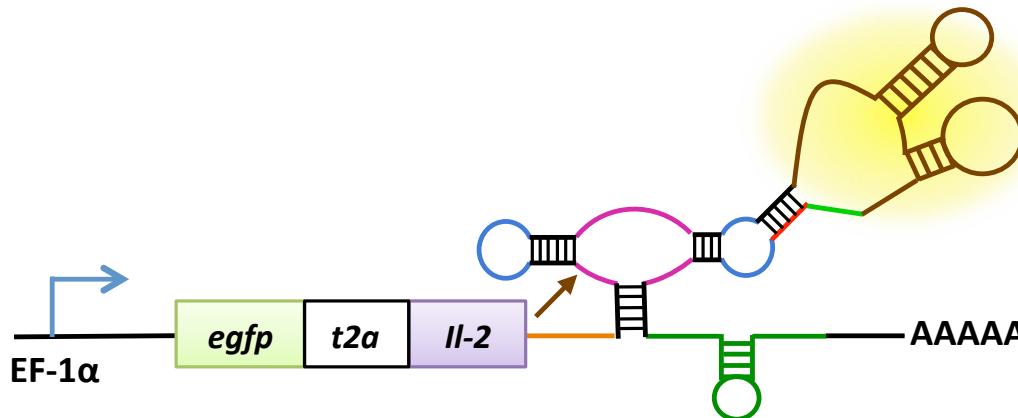
# Stable cell lines show dynamic response to input



# RNA devices regulate T-cell proliferation *in vivo*



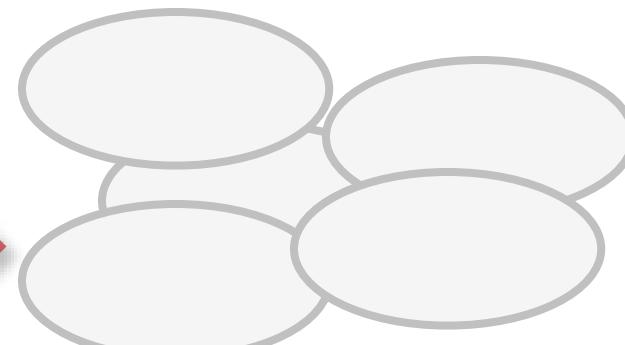
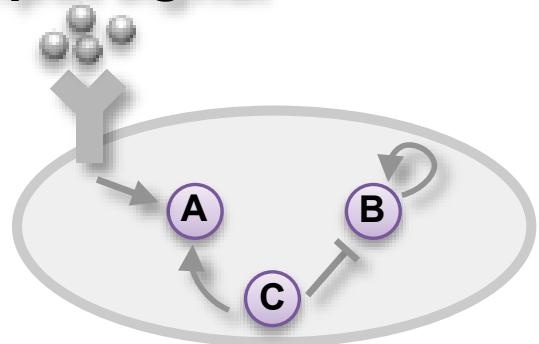
# Choice of drug-sensor pair affects device performance



# Moving between exogenous and embedded control

## Native Network

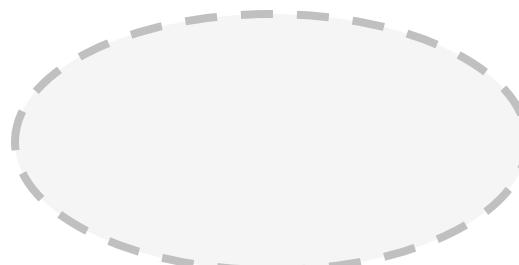
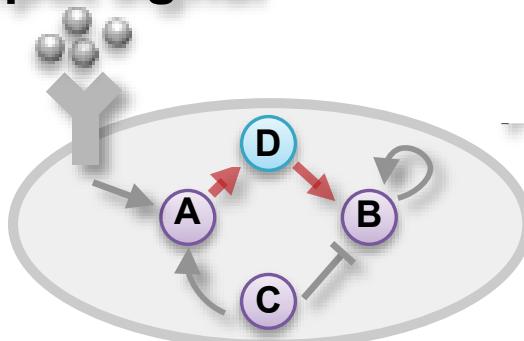
input signal



proliferation  
*fate 1*

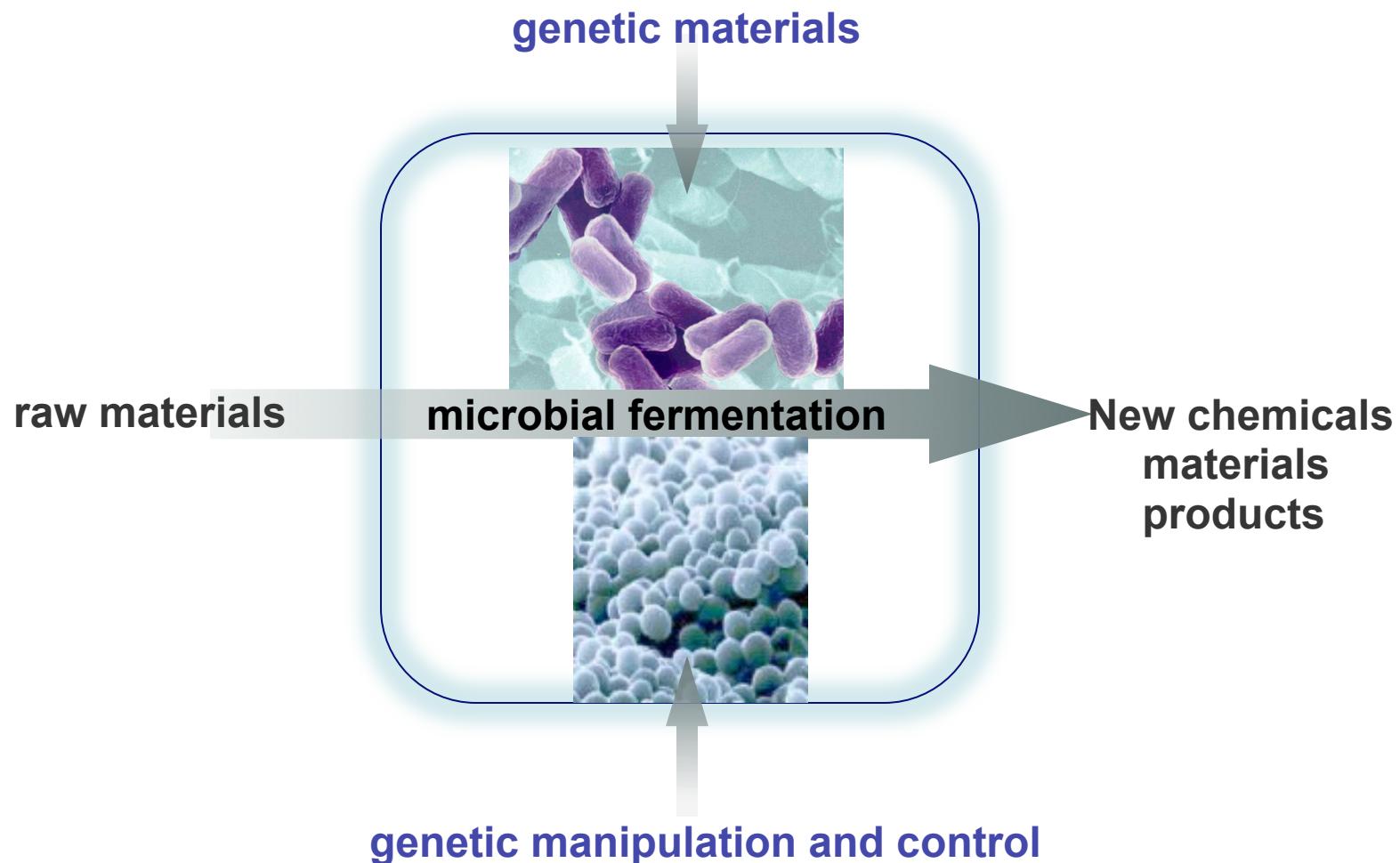
## Altered Network

input signal

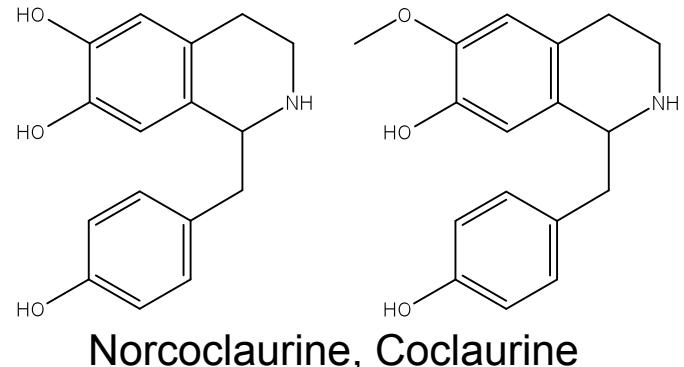
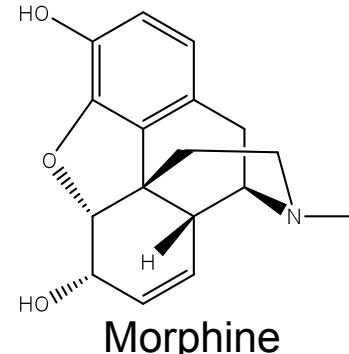
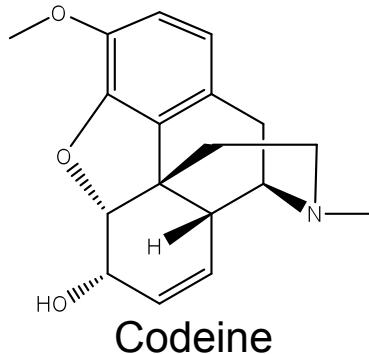


cell death  
*fate 2*

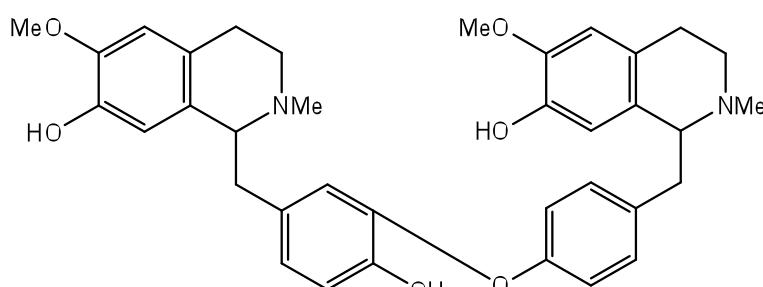
# Microbial factories



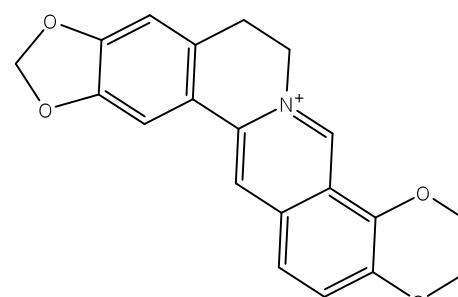
# Benzylisoquinoline Alkaloids (BIAs) are an important class of natural products



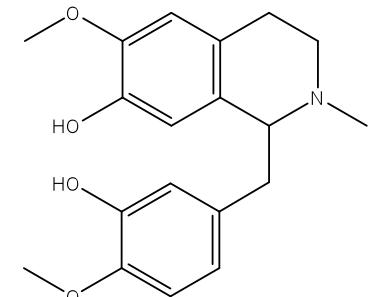
**Over 2,500 known structures**



Antimicrobial, anticancer

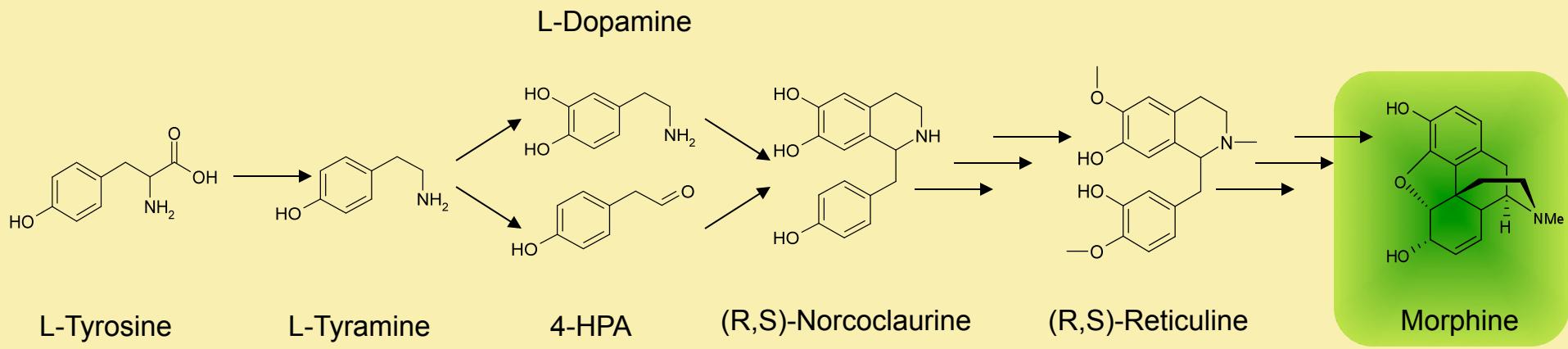


Antimicrobial, antioxidant



stimulates hair growth

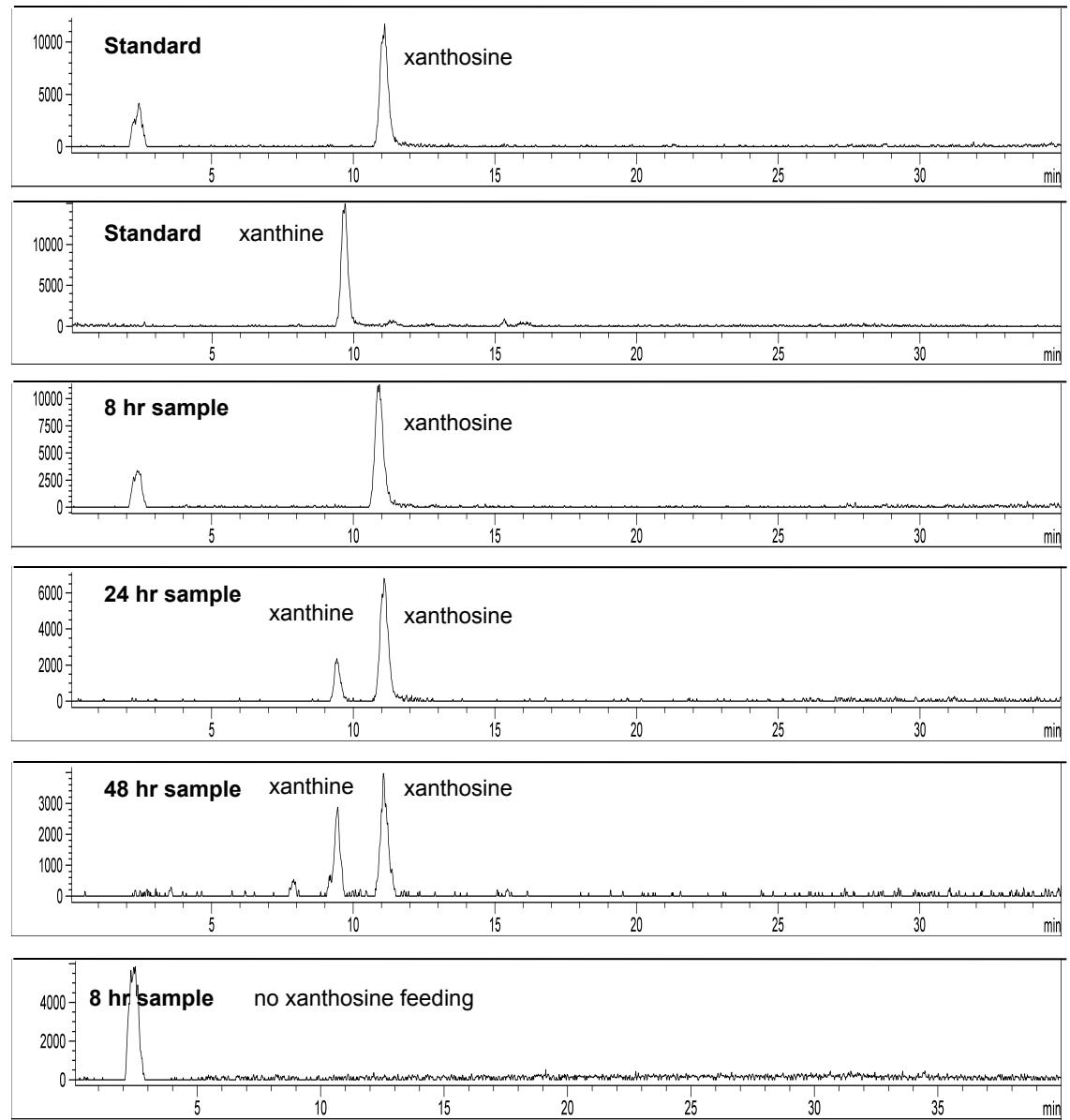
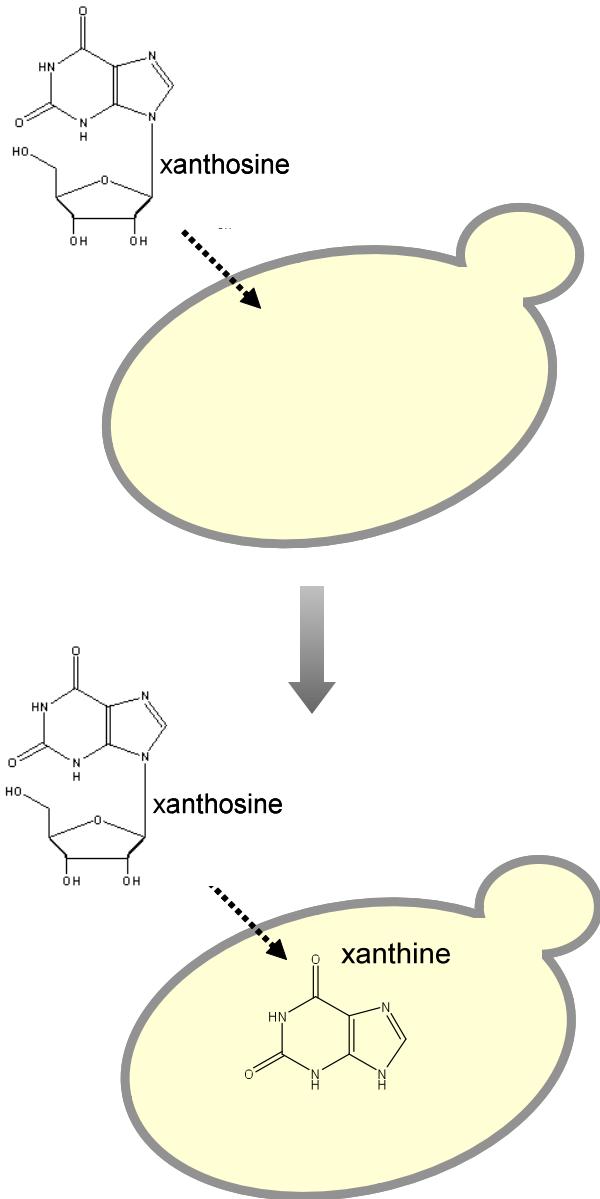
# Molecular tools for optimizing metabolite production



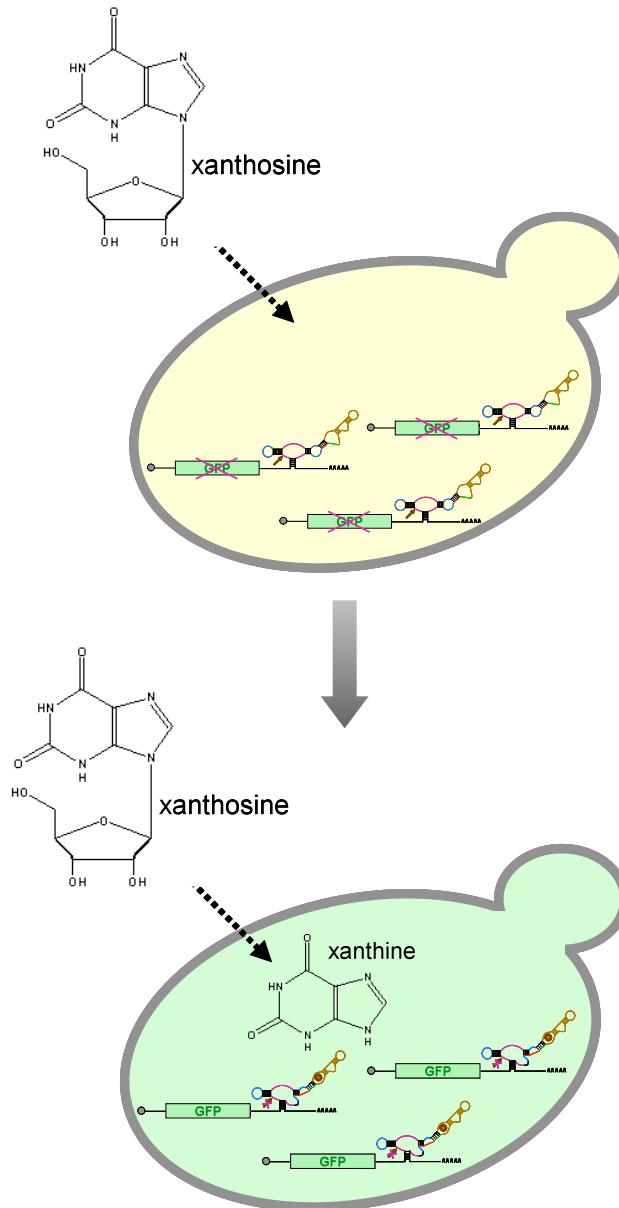
*Often need to screen through large libraries of pathway/enzyme variants – invasive / analytical procedures are too time and resource intensive*

***Engineer noninvasive sensors of metabolite concentrations in living cells***

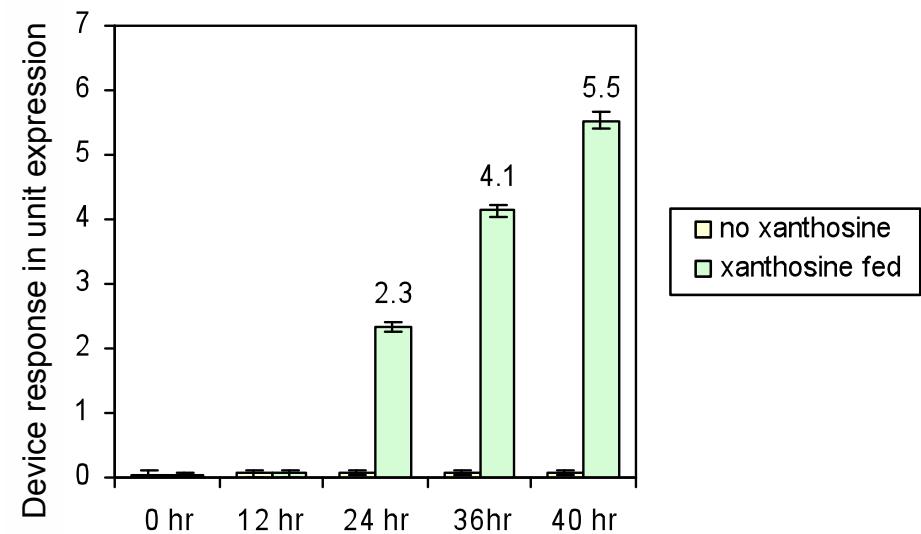
# Integrating RNA devices as noninvasive sensors of metabolite concentration



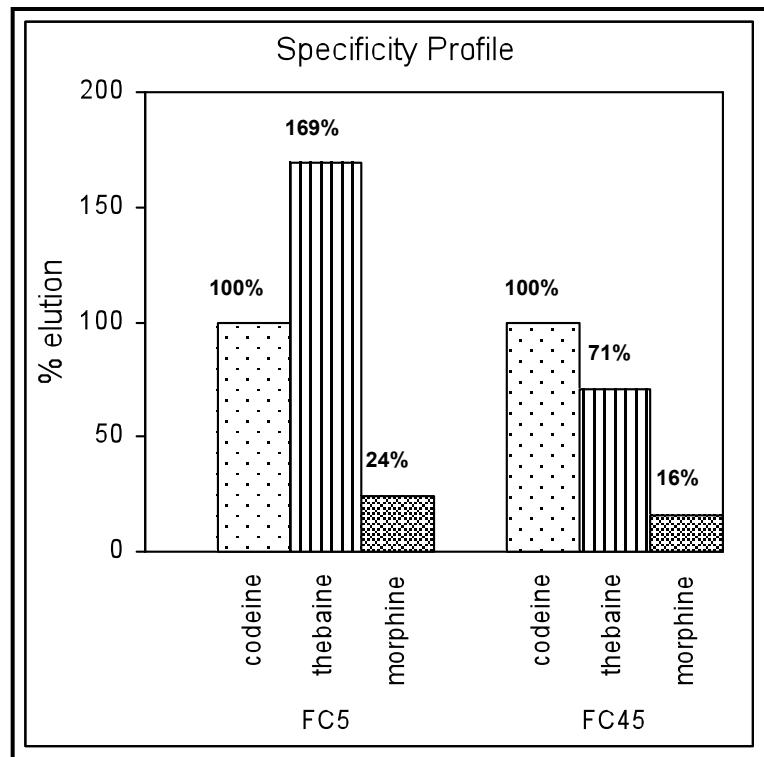
# RNA devices enable noninvasive sensing of metabolite concentration



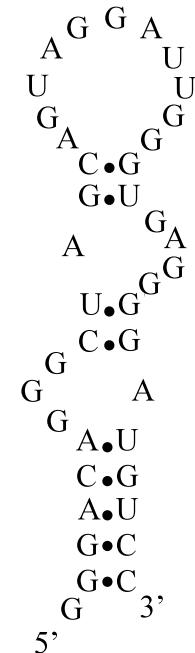
*Implementation of Buffer device  
as a real-time noninvasive  
xanthine sensor*



# RNA sensors generated to key BIAs

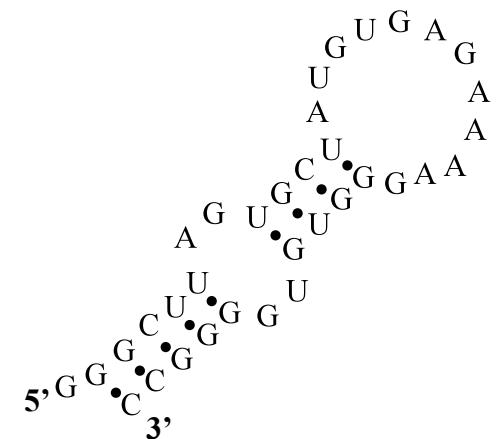


**FC5**

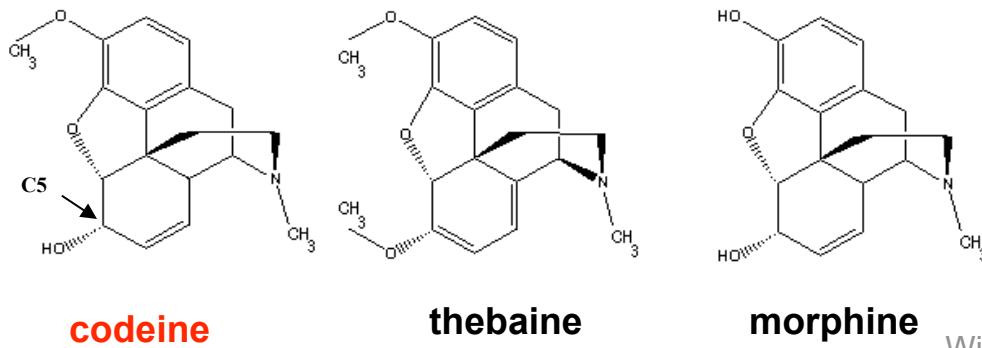


$$K_d (\text{FC5L}) = 4.55 \pm 0.14 \mu\text{M}$$

**FC45**



$$K_d (\text{FC45L}) = 2.59 \pm 0.09 \mu\text{M}$$



Win MN, Klein JS, Smolke CD. 2006. *Nuc Acids Res.* 34: 5670-82.

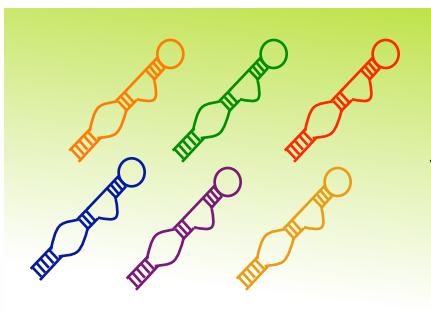
# Future directions: addressing the scalability challenge

refined sensor libraries

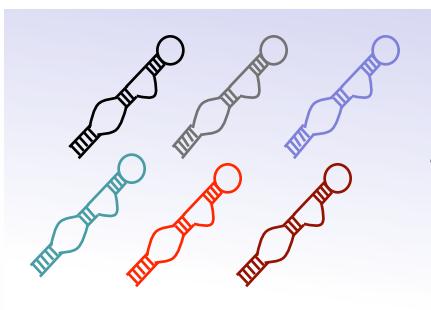
device platforms

applications

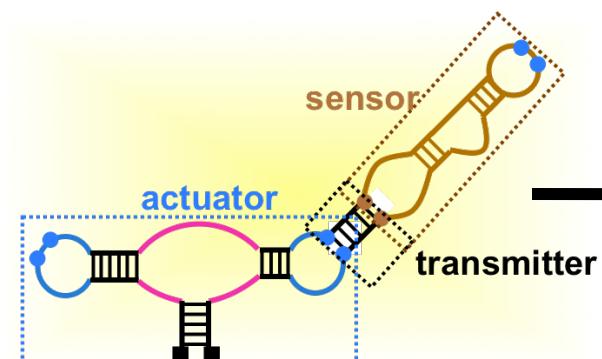
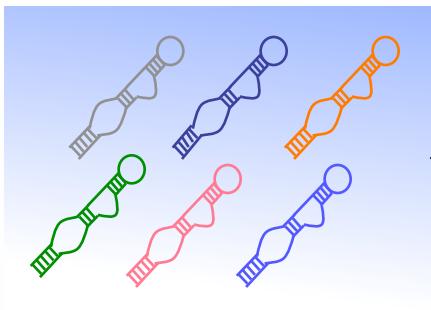
metabolites



disease biomarkers



exogenous chemicals



noninvasive  
diagnostics

intelligent  
therapeutics

bioprocessing /  
biosynthesis

agricultural  
biotechnology

***Optimization and development needed on selection  
and characterization strategies***

# Acknowledgements

## Current Lab Members

### Postdoctoral Researchers

Kate Thodey

### Graduate Researchers

Andrew Babiskin

Ryan Bloom

Arwen Brown

Andy Chang

Yvonne Chen

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Drew Kennedy

Joe Liang

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Michael Siddiqui

Isis Trenchard

Jay Vowles

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## Past Lab Members

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Chase Beisel

Stephanie Culler

Midori Greenwood-Goodwin

Kristy Hawkins

Kevin Hoff

Maung Nyan Win

## Collaborators

The Jensen Lab (City of Hope)

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